

1 **Effect of calcium soaps from garlic and *Salix babylonica* extracts on nematode loads,**
2 **nutrient intake and digestibility, nitrogen balance, and rumen fermentation kinetics in dairy**
3 **goats**

4 Einar Vargas-Bello-Pérez, Navid Ghavipanje, Teresa Torres-Gonzalez, Juan Carlos Angeles
5 Hernandez, Valente Velázquez-Ordoñez, Octavio Alonso Castelán Ortega, Lizbeth E. Robles
6 Jimenez, Sergio Roskof, Manuel Gonzalez-Ronquillo

7

8 **SUPPLEMENTARY FILE**

9

10

11

12 **Material & methods**

13 *Preparation of the extracts*

14 Every week, 4 L of Salix and Garlic extracts were prepared. For the preparation of Salix extract,
15 *S. babylonica* fresh leaves were randomly collected from mature trees during the summer and
16 autumn season (10 kg of fresh material). Leaves were chopped into 2–3 cm lengths and
17 immediately extracted soaking leaf material with water (1 g leaf per 8 ml of water). Plant material
18 was soaked and incubated at 25 °C for 72 h in closed jars of 5 L. After incubation, jars were
19 heated at 39 °C for 1 h and then filtered. Filtrated material was stored at 4 °C until its use. For
20 garlic, 10 kg of fresh material was obtained from a local supermarket. Both materials from Salix
21 and Garlic were ground at 2mm in a blender using 500 g from each one and then 1 kg of
22 safflower oil (*Carthamus tintorius*) was added, allowing the mixtures to stand for a period of 30
23 days in amber bottles for later use.

24 *Saponification process*

25 After a 30-day period, the process of fat saponification was performed following the double
26 decomposition method of Jenkins and Palmquist (1984), using safflower oil as control and the
27 Garlic and Salix extracts. Calcium soaps from safflower oil were chosen as control because they
28 have shown to have more energy density than those from with rapeseed, corn, soybean, and
29 sesame oils (Romero-Davila et al. 2017).

30 For calcium soaps preparation, in each of the three treatments, 1L of safflower oil, 250 g of
31 calcium chloride, 50 grams of calcium carbonate, 300 ml of alcohol 96%, 1370 g sodium
32 hydroxide at 30%, and 500 g of fresh leaves of Salix or Garlic was used. All the ingredients were
33 mixed until obtaining a homogeneous paste; the mixture was placed in glass containers in a

34 forced air oven at 60 °C for 48 hours to dehydrate each of the samples. This process was carried
35 out separately for each treatment.

36 *Rumen fermentation kinetics*

37 *In vitro* gas production (GP) test was performed according to Theodorou *et al.* (1994). Briefly,
38 800 mg of DM of each diet was placed in 125 mL glass flasks and 90 mL of buffer solution and
39 10 mL of rumen fluid were added to them. The buffer solution was prepared according to the
40 technique of Menke and Steingass (1988), in which 0.800 g DM of each ingredient and each diet
41 mixture were incubated in glass flasks of 125 ml, to which 90 ml of buffer solution and 10 ml of
42 rumen fluid were added, to make three bottles per sample. Three incubation runs were carried out
43 as replicates, in each incubation run, three flasks per treatment were used, a set of appropriate
44 blanks were included to adjust for the potential effects of other soluble materials in the extracts on
45 overall gas production and for readings correction of substrates, including bottles from self-
46 fermentation of rumen inoculum. Rumen fluid (300 ml per animal) was collected from the three-
47 rumen cannulated lactating goats (40 ± 3 kg of body weight, means ± SD) into a pre-warmed
48 thermos flask, and then filtered and flushed with CO₂. GP was measured using a pressure
49 transducer (Delta Model HD 8804, Italy) at 6, 12, 24, 48, 72, and 96 hr and a set of blanks were
50 included. GP was registered using a pressure transducer (Delta Model HD 8804, Italy). After
51 incubations, samples were filtered and dried for 48 h at 65 °C for dry matter disappearance
52 (DMd) determination. For relative gas production (RGP, mL gas/g DMd) determination, gas
53 production at 96 h was correlated with DMd. Kinetic parameters GP were estimated through an
54 iterative procedure of non-linear regression analysis (PROC NLIN, SAS Institute Inc., Cary, NC)
55 according to Krishnamoorthy *et al.* (1995):

$$56 \text{ GP} = B (1 - e^{-C(t-l)})$$

57 Where GP is the volume of gas (ml gas / g DM) at time t; B is the asymptotic GP (ml/gDM); C is
58 the fractional rate of GP (g/h), and l (h) is the discrete lag time prior to gas production.

59 After the *in vitro* incubation period, the samples were filtered and dried (48 h, 60 °C) to measure
60 the dry matter disappearance (DMD96h), and gas yield production at 24 h (GY24) was
61 determined. The volume of gas (ml gas/g DM) produced after 24 h of incubation was calculated
62 by dividing the amount of DMD (g):

$$63 \text{ Gas production (GY24)} = \text{ml gas 24h} / \text{g DMD96h}$$

64 The gas production (GP) at 96 h was correlated with DM disappeared to produce relative gas
65 yield (RGY; ml gas/g DMd) (González Ronquillo *et al.* 1998).

$$66 \text{ RGY} = \text{ml gas 96h} / \text{g DMd96h}$$

67 Short chain fatty acids concentration (SCFA) was calculated according to Getachew *et al.* (2002)
68 as: SCFA (mmol/200 mg DM) = 0.0222 GP - 0.00425. Where: GP is the 24 h net gas production
69 (ml/200 mg DM).

70 Microbial biomass production (MCP) was calculated according to Blümmel et al. (1997) as: MP
71 (mg/g DM) = mg IVDMD - (ml gas × 2.2 mg/ml). Where 2.2 mg/ml is a stoichiometric factor,
72 which expresses mg of C, H and O required for the SCFA gas associated with production of one
73 ml of gas (Blümmel et al. 1997).

74 *Chemical analysis for diets, feces, and urine*

75 To determine the DM content of feed, refusals and feces, samples were dried in a forced air oven
76 (60°C, 48 h), and subsequently ground in a Wiley mill 3mm Ø (Arthur H. Thomas, Philadelphia).
77 Organic matter, ether extract, and crude protein (N × 6.25) contents were determined using
78 standard procedures (AOAC 1990). Neutral detergent fiber (NDF) and acid detergent fiber (ADF)
79 were determined using the ANKOM technique (Van Soest et al. 1991) with alpha amylase and
80 uncorrected for ash.

81 **References**

82 **Jenkins TC and Palmquist DL** (1984) Effect of fatty acids or calcium soaps on rumen and total
83 nutrient digestibility of dairy rations. *J Dairy Sci* 67: 978–986. [https://doi.org/10.3168/jds.S0022-0302\(84\)81396-X](https://doi.org/10.3168/jds.S0022-0302(84)81396-X)

85 **Romero-Davila A, Herrera-Corredor C, Calderón-Aranda JA, Buendía-Rodríguez G,**
86 **Pescador Salas N and González-Ronquillo M** (2017) Uso de jabones de calcio como
87 alternativa de suplementación en rumiantes en épocas se sequia en zonas áridas y
88 semiáridas In: *Sustentabilidad agropecuaria ; experiencias de investigación para el*
89 *desarrollo agropecuario, forestal y rural*. Ciudad de México: Colofón. Estado de
90 México: Universidad Autónoma del Estado de México. 155-168. ISBN:
91 9786078583012

92 **Theodorou MK, Williams BA, Dhanoa MS, McAllan AB and France J** (1994) A simple gas
93 production method using a pressure transducer to determine the fermentation kinetics
94 of ruminant feeds. *Anim Feed Sci Technol* 48: 185–197. [https://doi.org/10.1016/0377-](https://doi.org/10.1016/0377-8401(94)90171-6)
95 [8401\(94\)90171-6](https://doi.org/10.1016/0377-8401(94)90171-6)

96 **Menke KH and Steingass H** (1988) Estimation of the energetic feed value obtained from
97 chemical analysis and gas production using rumen fluid. *Anim Res Dev* 28: 7–55.

98 **Van Soest PJ, Robertson JB and Lewis BA** (1991) Methods for dietary fiber, neutral detergent
99 fiber and nonstarch polysaccharides in relation to animal nutrition. *Br J Nutr* 74: 3583-
100 3597. [https://doi.org/10.3168/jds.S0022-0302\(91\)78551-2](https://doi.org/10.3168/jds.S0022-0302(91)78551-2)

101 **Krishnamoorthy U, Soller H, Steingass H and Menke KH** (1995) Energy and protein
102 evaluation of tropical feedstuffs for whole tract and ruminal digestion by chemical

103 analysis and rumen inoculum studies in vitro. *Anim Feed Sci Technol* 52: 177–188.
104 <https://doi.org/10.14202/vetworld.2015.605-609>

Table S1. Chemical composition (g/kg DM) of ingredients used in experimental diets.

Item	Safflower calcium soap	Garlic calcium soap	Salix calcium soap	Alfalfa hay	Rapeseed	Sorghum grain	SBM	Wheat bran	Corn stover
Dry matter	886	874	860	930	920	898	880	874	917
Organic matter	607	628	463	900	930	980	930	955	945
Crude protein	2	9	4	170	360	100	460	160	50
RDP	2	4	2	119	252	70	322	112	35
Fat	701	615	533	30	32	35	20	40	20
NDF	14	15	18	449	305	430	134	487	722
ADF	8	10	12	254	152	58	92	122	410
ADL	3	2	7	128	98	44.5	35	68	90
ME, Mj/Kg DM	25.0	23.0	21.0	10.0	12.1	13.5	13.0	11.0	7.8

SBM, soybean meal; RDP, Rumen degradable protein; NDF, Neutral Detergent Fiber; ADF, Acid Detergent Fiber; ADL, AcidDetergent. Lignin; ME, Metabolizable energy (MJ/kg DM).

Table S2. Ingredients and chemical composition (g/kg DM) from control, Garlic and Salix diets

Item	Treatments ¹		
	Control	Garlic	Salix
Ingredients (g/kg DM)			
Alfalfa hay	600	600	600
Corn stover	60	60	60
Sorghum grain	121	121	121
Soybean meal	40	40	40
Rapeseed	40	40	40
Wheat bran	40	40	40
Calcium soaps	65	65	65
Vitamin and mineral premix ²	33	33	33
Chemical composition			
Dry matter, DM	939	938	937
Organic matter, OM	878	867	884
Crude protein, CP	143	136	136
Neutral detergent fiber	426	412	440
Acid detergent fiber, ADF	224	216	228
Acid detergent lignin, ADL	97	72	62.5
Ether extract, EE	120	118	128

¹ Diets supplemented with 65 g/kg DM of calcium soaps from either safflower (Control), garlic extract (Garlic), or *Salix babylonica* extract (Salix), respectively.

²Containing in 1.0 kg DM the following: 25 mg of antioxidant, 4.5 g of calcium carbonate, 6 g of salt, 30 g of ionophore, 50 g of zinc oxide, 6 g of sodium bicarbonate, 6 g of copper sulphate, 20 g of ferrous sulphate, 125 g of sodium sulphate, 18,000 IU of vitamin E, 3,000,000 UI of vitamin A, 3,750,000 IU of vitamin D, 140 g of potassium chloride, 0.500 g of EDD. I ethylene-dynamine, 0.090 g of cobalt carbonate, 500 mg of magnesium oxide, 36 g of manganese oxide and 0.090 g of selenium.

Table S3. *In vitro* rumen gas kinetics (ml gas/g DM) and fermentation profile of goats fed with calcium soaps of Garlic and Salix extracts.

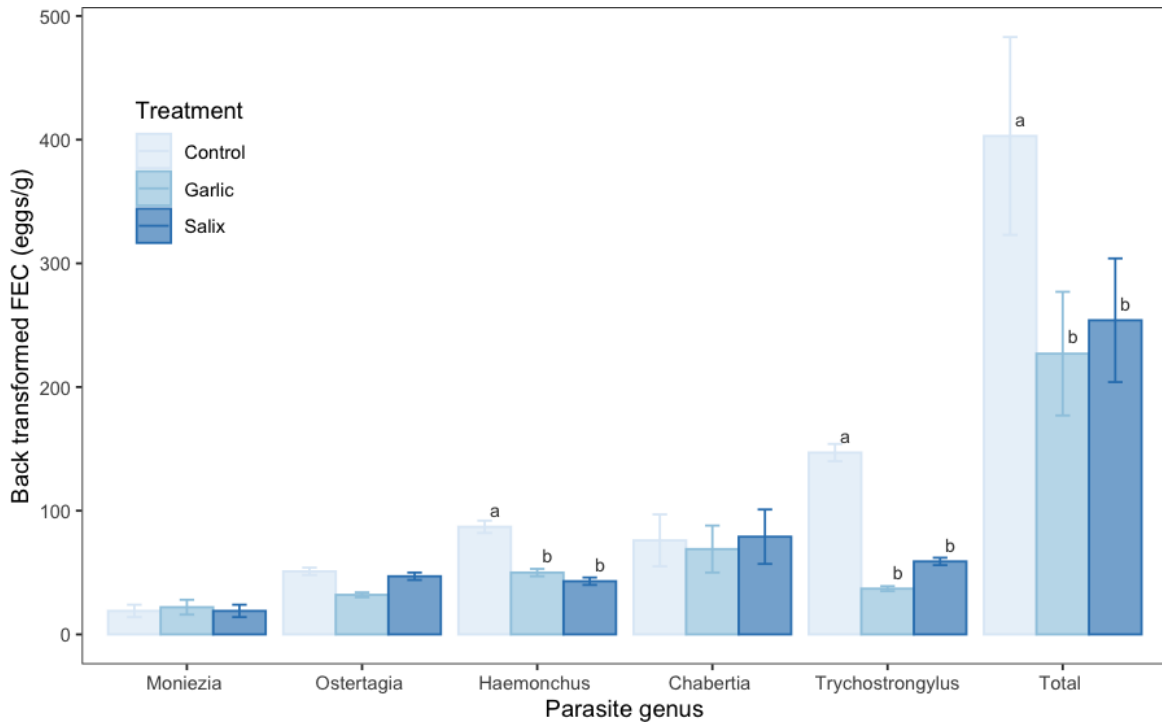
Parameters ¹	Treatments ²			SEM	P-value
	Control	Garlic	Salix		
A	179	191	183	9.43	0.250
B	0.04	0.04	0.42	0.003	0.333
Lag time	2.39	2.23	2.00	0.29	0.237
Gas production, ml gas/g DM					
6 h	18.9	21.3	24.2	3.21	0.120
12 h	67.9	67.4	72.3	6.47	0.521
24 h	104	109	112	6.28	0.257
48 h	146	155	152	7.74	0.314
96 h	177	187	183	8.55	0.256
DMD96	585	587	582	15.0	0.897
RGP	302	319	315	15.9	0.340
GY24	178	186	192	13.1	0.361
SCA	0.30	0.30	0.30	0.002	0.356
MCP	555	557	550	16.8	0.835
ME, MJ/kg DM	14.0	13.6	13.9	0.34	0.224

^{a-c} Means with different superscript letters within a row are different ($P \leq 0.05$).

¹ A, total gas production (ml gas/g DM incubated); B, fermentation rate (h⁻¹); C, fermentation rate (h^{-1/2}); L, the initial delay before gas production begins (h); DMD96, DM degraded substrate (mg/g DM); RGP, relative gas production (mL gas/g DMD96); SCFAs, short chain fatty acids (mmol/g DM); GY24, gas yield at 24 h (mL gas/g DMD); MCP, microbial CP production (mg/g DM).

² Diets supplemented with 65 g/kg DM of calcium soaps from either safflower (Control), garlic extract (Garlic), or *Salix babylonica* extract (Salix), respectively

SEM = Standard error of the mean

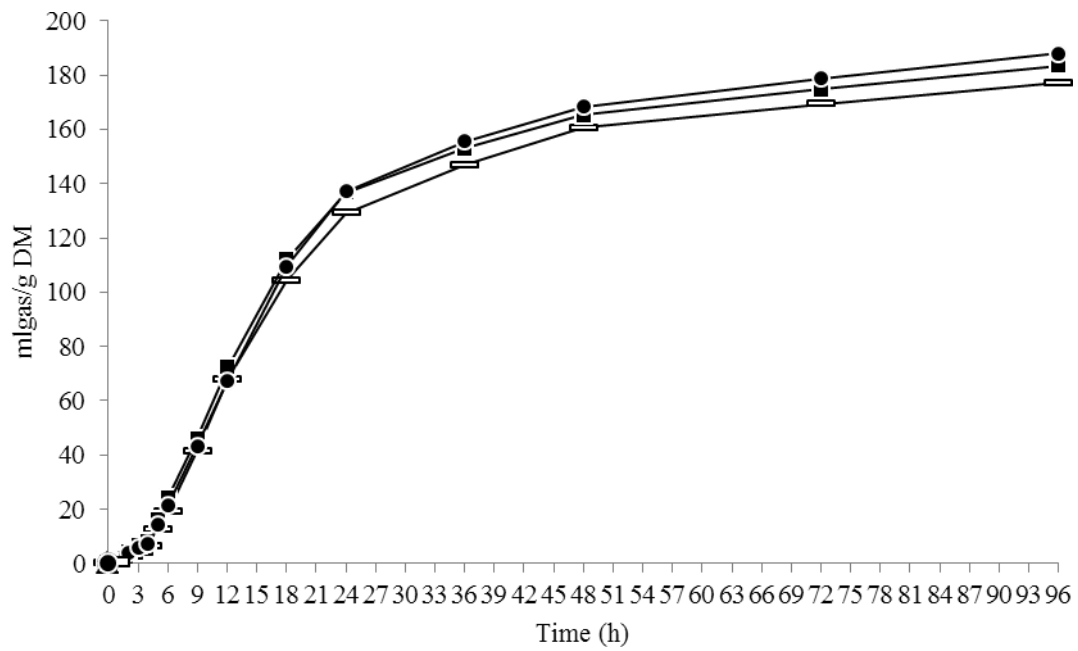


104

105

106

Figure S1. Back transformed FEC (egg/g) of according with parasite genus.



107

108 **Figure S2.** Cumulative gas production at 96 h (ml gas / g DM) of diets supplemented with
 109 calcium soaps of Safllower (□), Garlic (●), and Salix (■) extracts.