

**Valentin PARTULA\*, Mélanie DESCHASAUX-TANGUY\*, et al. Associations between untargeted plasma metabolomic signatures and gut microbiota composition in the *Milieu Intérieur* population of healthy adults**

**SUPPLEMENTARY MATERIAL**

**Supplementary Material and Method 1.** Gut microbiota sequencing protocol, Milieu Intérieur study, France, 2012.

**DNA EXTRACTION**

Bacterial DNA was extracted from stool samples using a Repeated Bead Beating Plus Column (RBB+C) method, as previously published (O'Cuiv et al. 2011; Yu et al. 2004). Cell lysis was achieved by bead beating in the presence of 4% (w/v) sodium dodecyl sulfate, 500mM sodium chloride, and 50mM ethylenediaminetetraacetic acid. Consequently to bead beating, the sodium dodecyl sulfate and most of the impurities were removed by precipitation with ammonium acetate. Nucleic acids were recovered by precipitation with isopropanol. Subsequent sequential digestions with RNase and proteinase K were used to remove RNA and proteins, respectively. Finally, genomic DNA was purified using QIAamp columns (Qiagen, Germantown, MD, USA); then recovered DNA was checked for quality and quantity using NanoDrop and Qubit together with Broad Range assay (ThermoFisher Scientific, Waltham, MA, USA).

**BARCODING POLYMERASE CHAIN REACTION (PCR)**

Primers targeting the V3-V4 hypervariable regions of the 16S rRNA gene were used for the barcoding PCR (forward primer: CCTACGGRAGGCAGCAG; reverse primer: GGACTACHVGGGTWTCTAAT) (Kozich et al. 2013). PCR mix was made up of 18µL of AccuPrime™ Pfx SuperMix (Invitrogen-12344-040, ThermoFisher Scientific, Waltham, MA, USA), 0.5µL of both V3-340F and V4-806R primers (0.2µM), and 1µL of DNA (10ng). PCR cycle sequence was: 2min-95°C; 30 cycles of 20sec-95°C; 15sec-55°C; 5min-72°C; and 10min-72°C. PCR reaction was quantified using the Qubit BR DNA assay kit (ThermoFisher Scientific, Waltham, MA). Equal amounts of each PCR reaction were pooled and thoroughly mixed. The amplicon pool was then loaded on a 1.5%-agarose gel and electrophoresis was

run for 45min at 100V. The amplicon band corresponding to the 16S rDNA V3-V4 was extracted and purified using Qiaquick gel extraction kit (Qiagen-28704, Qiagen, Germantown, MD, USA). The amplicon quantity and integrity were checked on agarose gel electrophoresis and Qubit BR DNA assay kit (ThermoFisher Scientific, Waltham, MA, USA). Amplicon was stored at -20°C.

## **SEQUENCING**

16S rDNA amplicon was sequenced at Institut Curie NGS platform on a MiSeq machine (Illumina, San Diego, CA, USA) using the 2\*300 bp V3 kit.

### *Bioinformatics pipeline and computation of microbial diversity indexes*

Pipelines sickle (Joshi et al. 2011), SPAdes (Bankevich et al. 2012), and PEAR (Zhang et al. 2014) were used to trim, error correct, and merge reads, respectively. vsearch (Rognes et al. 2016) was used to cluster reads at 97% of identity, in order to generate Operational Taxonomic Units (OTU). UCHIME (Edgar et al. 2011) was used to identify chimeric OTUs which were discarded from further analysis. RDP SeqMatch and reference database RDP Release 11.3 were used to determine taxonomy of representative OTU sequences. These were then aligned using ssu-align (Nawrocki 2009). Fasttree2 (Price et al. 2010) was used to infer the phylogenetic tree from the OTUs multiple alignments.

R package *vegan* was used to generate microbial diversity indicators – namely Simpson's index, observed richness, and Chao1 richness estimate ( $\alpha$ -diversity) and Bray-Curtis dissimilarity ( $\beta$ -diversity) – from the non-rarefied OTU count table.

## **LITERATURE CITED**

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**Supplementary Material and Method 2.** Metabolomic analysis protocol, Milieu Intérieur study, France, 2012.

### **NMR SAMPLE PREPARATION**

350 $\mu$ L of plasma (thawed at room temperature) were mixed with 250 $\mu$ L of buffer solution (10mmol.L<sup>-1</sup> phosphate buffer saline/5mmol.L<sup>-1</sup> NaN<sub>3</sub> in D<sub>2</sub>O). Mix was centrifuged (12,000g, 5min, 4°C) and supernatant placed into a NMR tube.

### **ONE-DIMENSIONAL NMR SPECTRA ACQUISITION**

For each plasma sample, two complementary one-dimensional NMR sequences were acquired on a 500MHz-Bruker Avance III spectrometer (Bruker, Billerica, MA, USA) at 300K – namely the Nuclear Overhauser Effect Spectroscopy (NOESY) and the Carr-Purcell-Meiboom-Gill (CPMG) sequences. Both were used with 128 transients. The spectral window was 10 kHz. The water signal was suppressed by a presaturation sequence using low-power irradiation on the water-signal frequency during the relaxation delay.

### **POST-ACQUISITION SPECTRA PRE-PROCESSING**

A Fourier transform was applied with an exponential window function to produce a 0.3 Hz broadening line. A zero-order phase and a manual baseline correction were applied to each spectrum using Bruker's TopSpin 4.0.3. Spectra were aligned using the icoshift algorithm (Savorani et al. 2010) and normalized using the probabilistic quotient normalization method (Dieterle et al. 2006). Finally, the grouped-batch profile calibration strategy (Fages et al. 2013) was used to remove variation presumably introduced by experimental procedures (i.e. batch-effect).

### **INTELLIGENT BUCKETING OF SPECTRA AND OBTAINING OF NMR VARIABLES**

After processing and calibration, each spectrum was sliced into bins with pre-defined limits so as to maximize recovery of peak entities. This "intelligent bucketing" procedure (Sousa et al. 2013) mitigates spoiling of peak integrity compared to bucketing with steady 0.001 ppm-intervals. Then, the buckets were scaled to the total summed integrals for each spectrum,

and the integrals of peak entities were calculated to obtain continuous NMR variables that were used in further statistical analyses.

### **NMR SIGNALS ASSIGNMENTS**

NMR signals were assigned using ad hoc literature (Nicholson et al. 1995). CHENOMX software (CHENOMX, Edmonton, Alberta, Canada) and Human Metabolome Database (Wishart et al. 2013) were used to confirm the assignments using the medium spectrum (Nicholson et al. 1995). After the statistical analysis, the assignments of the buckets that had an association with the gut microbia were newly carefully looked and confirmed to be unambiguously assigned by gathering information from S-TOCSY, additional two-dimension NMR experiments (recorded on random samples) such as 1H–1H Total Correlated Spectroscopy (TOCSY) and J-resolved experiments.

### **LITERATURE CITED**

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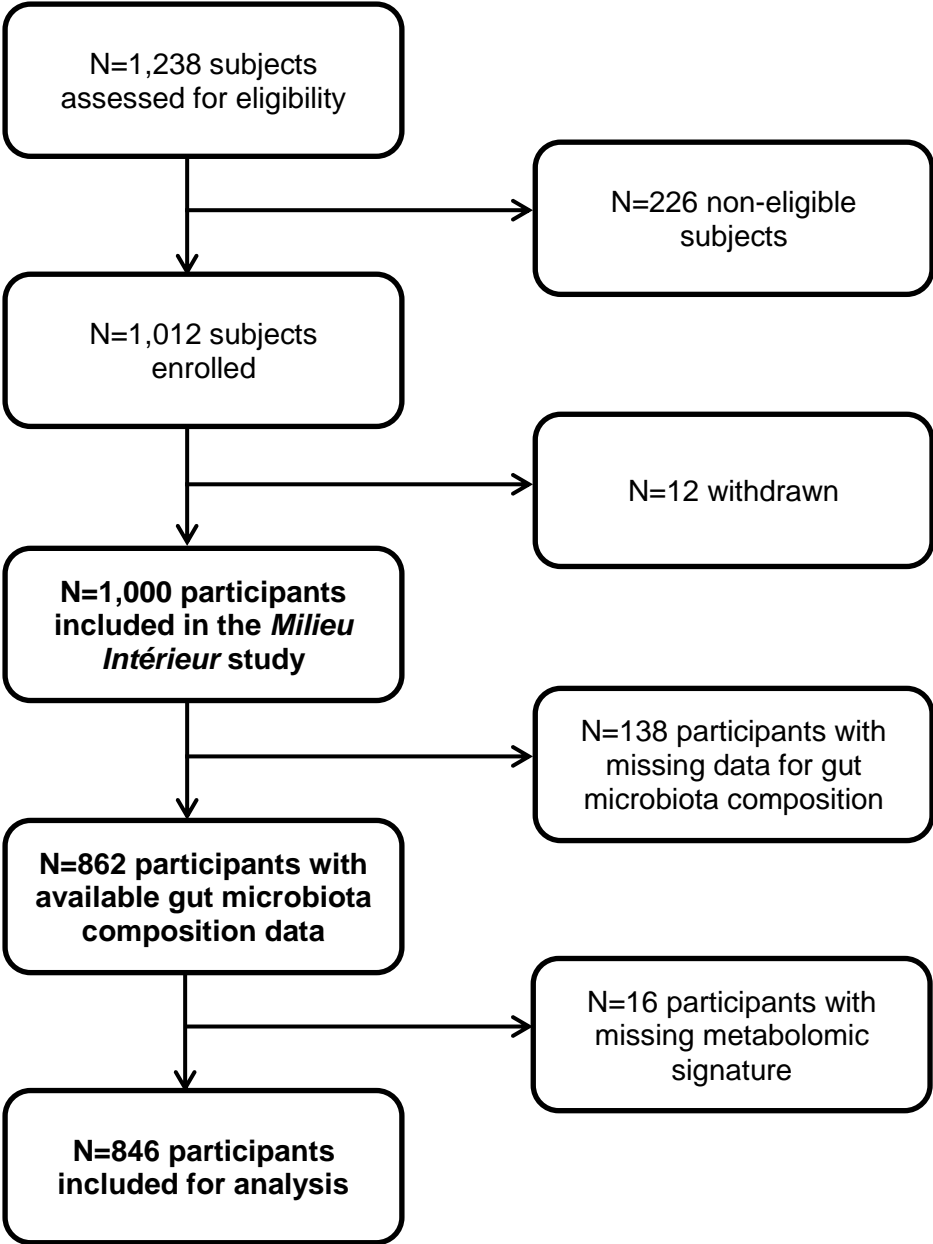
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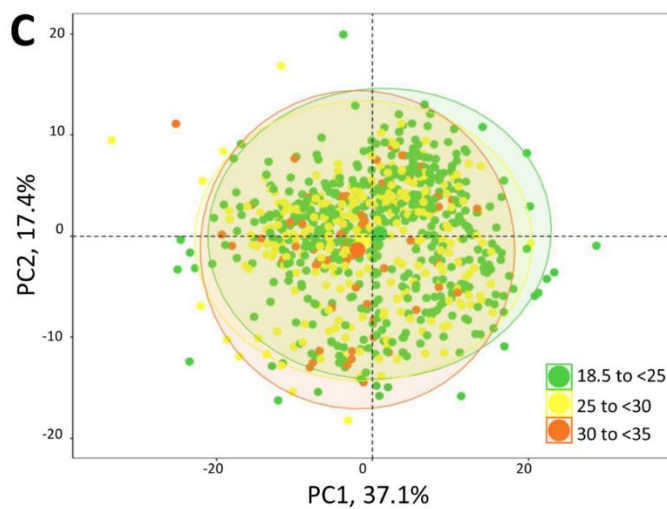
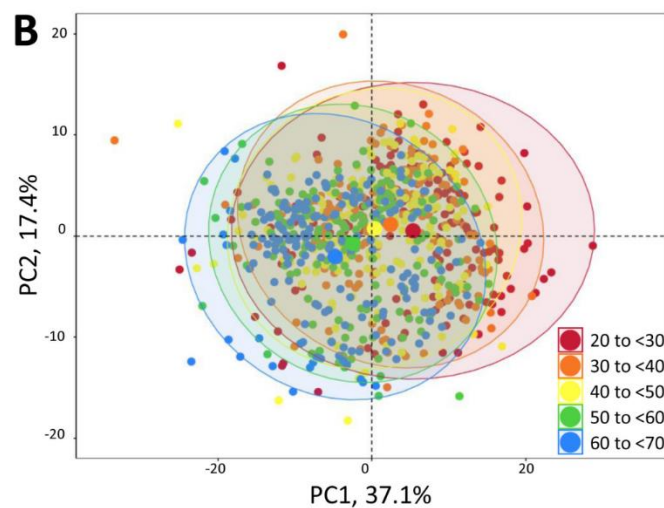
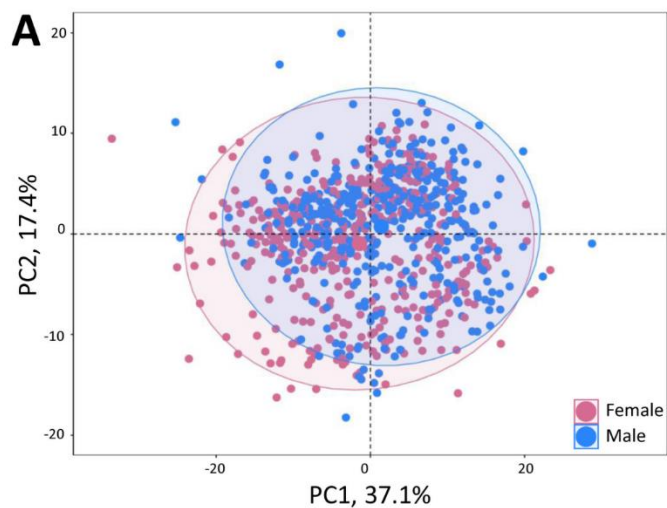
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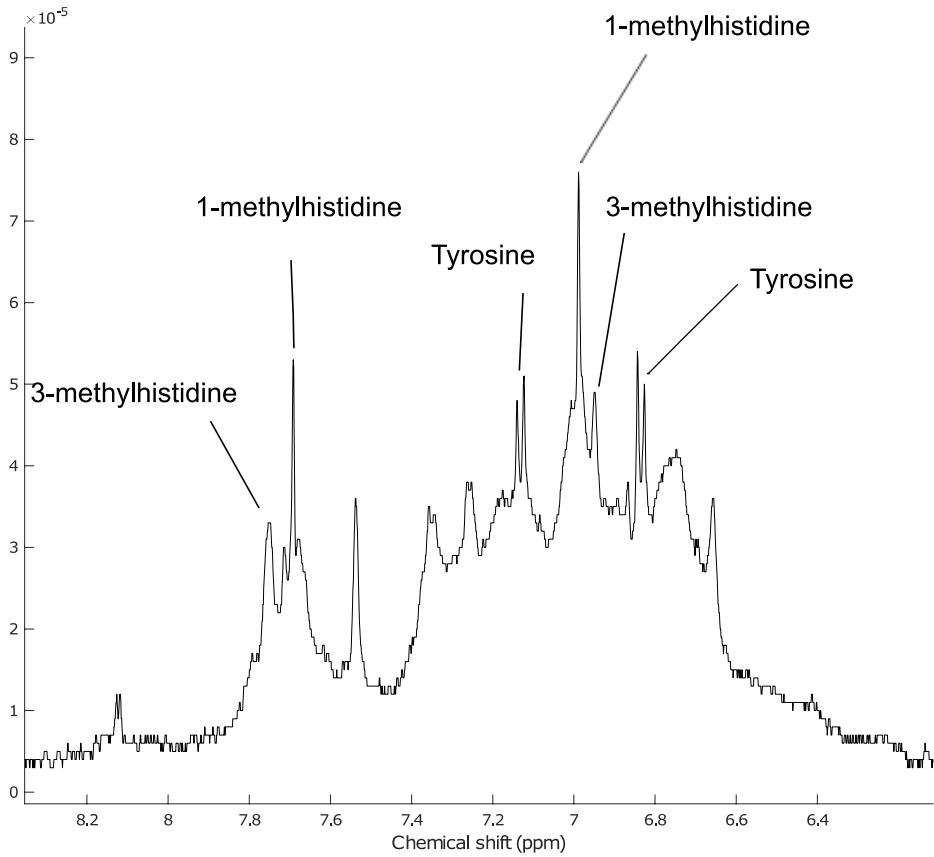
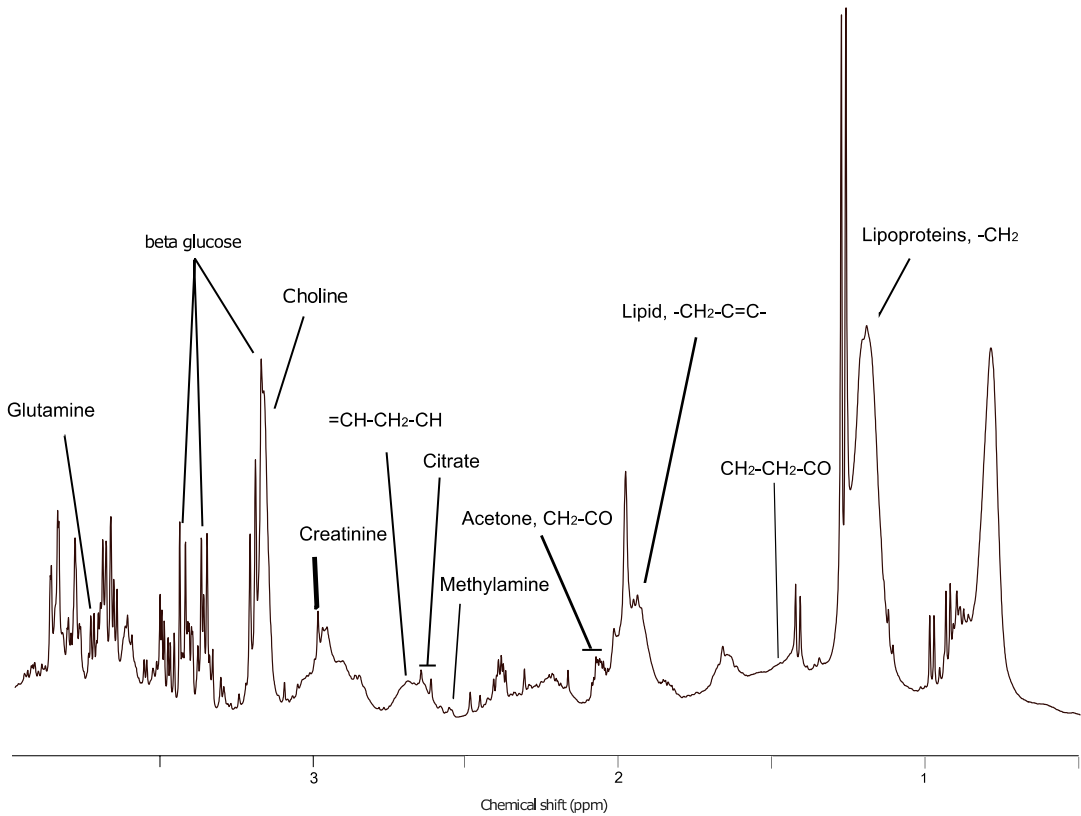
Supplementary Figure 1. Participant flow chart, *Milieu Intérieur* study, France, 2012.



**Supplementary Figure 2.** Interindividual variation in metabolomic signatures represented by principal components (PCA) analysis of the NOESY1D metabolomic dataset, *Milieu Intérieur* study, France, 2012 (N=846). Each point represents an individual from the study sample. PCA was obtained via the *PCA* function (package *FactoMineR*), and plotted and color-coded based on sex (A), age (B), and BMI (C) via the *fviz\_pca\_ind* function (package *FactoExtra*). Concentration ellipses (95%) are shown. Percentages on the axes represent the proportion of variation explained by the two first components of the PCA.



**Supplementary Figure 3.** Medium NMR spectrum. The buckets that have been highlighted in the manuscript have been assigned on the spectrum.





**Supplementary Table 1.** “Core microbiota” (i.e. genera shared by at least 95% of the study population), *Milieu Intérieur* study, France, 2012 (N=846).

Phylum  Class  Order  Family  Genus <sup>1</sup>	Coverage (%) <sup>2</sup>
Firmicutes  Clostridia  Clostridiales  Ruminococcaceae  Flavonifractor <sup>3</sup>	100
Firmicutes  Clostridia  Clostridiales  Lachnospiraceae  Clostridium XIVa <sup>2</sup>	100
Firmicutes  Clostridia  Clostridiales  Lachnospiraceae  Lachnospiraceae incertae sedis <sup>2</sup>	100
Firmicutes  Clostridia  Clostridiales  Lachnospiraceae  Blautia <sup>2</sup>	100
Firmicutes  Clostridia  Clostridiales  Ruminococcaceae  Clostridium IV <sup>2</sup>	100
Bacteroidetes  Bacteroidia  Bacteroidales  Bacteroidaceae  Bacteroides <sup>2</sup>	99.9
Firmicutes  Clostridia  Clostridiales  Eubacteriaceae  Eubacterium	99.9
Firmicutes  Clostridia  Clostridiales  Ruminococcaceae  Acetivibrio	99.9
Firmicutes  Clostridia  Clostridiales  Ruminococcaceae  Oscillibacter <sup>3</sup>	99.8
Firmicutes  Clostridia  Clostridiales  Lachnospiraceae  Coprococcus <sup>2</sup>	99.8
Firmicutes  Clostridia  Clostridiales  Lachnospiraceae  Ruminococcus II	99.8
Firmicutes  Clostridia  Clostridiales  Ruminococcaceae  Gemmiger <sup>3</sup>	99.7
Firmicutes  Clostridia  Clostridiales  Lachnospiraceae  Roseburia <sup>2</sup>	99.5
Firmicutes  Clostridia  Clostridiales  Ruminococcaceae  Faecalibacterium <sup>2</sup>	99.4
Firmicutes  Clostridia  Clostridiales  Ruminococcaceae  Intestinimonas	98.7
Firmicutes  Clostridia  Clostridiales  Lachnospiraceae  Fusicatenibacter	98.7
Bacteroidetes  Bacteroidia  Bacteroidales  Rikenellaceae  Alistipes <sup>2</sup>	98.5
Bacteroidetes  Bacteroidia  Bacteroidales  Porphyromonadaceae  Parabacteroides <sup>2</sup>	98.2
Firmicutes  Clostridia  Clostridiales  Lachnospiraceae  Dorea <sup>2</sup>	98.1
Firmicutes  Clostridia  Clostridiales  Ruminococcaceae  Clostridium III	98.1
Firmicutes  Clostridia  Clostridiales  Ruminococcaceae  Ruminococcus <sup>3</sup>	96.9
Firmicutes  Clostridia  Clostridiales  Lachnospiraceae  Eisenbergiella	95.7

<sup>1</sup> Genera shared with the “Western” core described throughout 4,000 samples from 4,000 European or North American individuals (\*\*) or found in the 95% core of at least one cohort (\*), as reported by Falony et al. (Falony G, et al. Population-level analysis of gut microbiome variation. *Science*. 2016. 352 (6285): 560-564).

<sup>2</sup> Prevalence of genus in the study population.

**Supplementary Table 2.** Relative abundance, taxonomical assignment and coverage of the detected genera and species. Milieu Intérieur study, France, 2012 (N=846)

*Provided as an Excel file*

**Supplementary Table 3.** Associations between covariates and Bray-Curtis dissimilarity, using univariate PERMANOVAs (999 permutations), *Milieu Intérieur* study, France, 2012 (N=846).

Covariate	Bray-Curtis dissimilarity	
	R <sup>2</sup>	p-value <sup>1</sup>
Age	0.005	0.001
Sex	0.006	0.001
BMI	0.001	0.4
Smoking status	0.004	0.008
Physical activity	0.001	0.5
Sequencing depth	0.07	0.001

<sup>1</sup> p-value for univariate PERMANOVA, 999 permutations, computed using the *adonis* function of the package *vegan*.

**Supplementary Table 4.** Description of the 202 CPMG continuous variables used in statistical analyses, *Milieu Intérieur* study, France, 2012 (N=846).

Spectral region (ppm)	Metabolite assignment	Mean	SD
0.527-0.567		0.00100	0.00007
0.567-0.607		0.00133	0.00008
0.607-0.647		0.00166	0.00007
0.647-0.713	CH <sub>3</sub> cholesterol	0.00344	0.00017
0.713-0.727	CH <sub>3</sub> cholesterol	0.00112	0.00006
0.727-0.767		0.00754	0.00044
0.767-0.807	CH <sub>3</sub> lipoproteins	0.03093	0.00287
0.807-0.911	CH <sub>3</sub> lipoproteins	0.04315	0.00242
0.911-0.937	Leucine, valine, isoleucine	0.00461	0.00024
0.937-0.983	Leucine, valine, isoleucine	0.00537	0.00028
0.983-1.019	Leucine, valine, isoleucine	0.00339	0.00017
1.019-1.053	Valine	0.00214	0.00011
0.911-0.97	Leucine	0.00801	0.00040
0.97-1.0	Valine	0.00391	0.00021
1.0-1.021	Isoleucine	0.00141	0.00008
1.021-1.051		0.00188	0.00010
1.051-1.053		0.00019	0.00001
1.053-1.081	Isobutyrate	0.00265	0.00010
1.081-1.088		0.00116	0.00004
1.088-1.127		0.10672	0.00504
1.127-1.167	Propylene glycol	0.06318	0.00253
1.127-1.145	Propylene glycol	0.05066	0.00237
1.145-1.167		0.01345	0.00074
1.167-1.207	CH <sub>2</sub> lipoproteins	0.04104	0.00335
1.207-1.311	CH <sub>2</sub> lipoproteins	0.13277	0.01211
1.311-1.351	Lactate threonine/CH <sub>2</sub> lipoproteins	0.00684	0.00040
1.351-1.391	CH <sub>2</sub> lipoproteins	0.00396	0.00019
1.391-1.431	Lysine	0.00677	0.00036
1.431-1.437		0.00110	0.00006
1.437-1.447		0.00129	0.00007
1.447-1.521	Alanine	0.01054	0.00063
1.521-1.527	CH <sub>2</sub> CH <sub>2</sub> COOC	0.00148	0.00015
1.527-1.631	CH <sub>2</sub> CH <sub>2</sub> COOC	0.01103	0.00049
1.631-1.636		0.00063	0.00004
1.636-1.647		0.00133	0.00009
1.647-1.751	Arginine lysine	0.00845	0.00060
1.647-1.707	Arginine	0.00604	0.00043
1.707-1.759	Lysine	0.00288	0.00021
1.759-1.791		0.00156	0.00011
1.751-1.791		0.00197	0.00014
1.791-1.821		0.00152	0.00010
1.821-1.849		0.00191	0.00013
1.849-1.89	Lysine acetate	0.00550	0.00020
1.89-1.927	Lysine acetate	0.00589	0.00032
1.849-1.899	Lysine	0.00648	0.00024

1.899-1.927	Acetate	0.00495	0.00028
1.927-1.967	Glycoproteins N-acetyl, CH <sub>2</sub> CH	0.01230	0.00078
1.967-2.063	Glycoproteins N-acetyl, CH <sub>2</sub> CH	0.02268	0.00102
1.927-2.017	CH <sub>2</sub> CH	0.02872	0.00128
2.017-2.063	Glycoproteins N-acetyl	0.00614	0.00033
2.063-2.094	Methionine	0.00360	0.00023
2.094-2.151	Methionine	0.00354	0.00017
2.151-2.166	Lipid -CH <sub>2</sub> =C=C-	0.00198	0.00022
2.166-2.167	Lipid -CH <sub>2</sub> =C=C-	0.00036	0.00004
2.167-2.264	Acetone CH <sub>2</sub> CO	0.01194	0.00068
2.264-2.311		0.00339	0.00022
2.264-2.277	Acetoacetate	0.00103	0.00006
2.277-2.311		0.00243	0.00017
2.311-2.339		0.00195	0.00013
2.311-2.315		0.00052	0.00004
2.315-2.354	Glutamate	0.00243	0.00016
2.354-2.372	Pyruvate	0.00124	0.00008
2.372-2.411		0.00387	0.00031
2.339-2.386	Pyruvate	0.00353	0.00025
2.386-2.471	Glutamine	0.00518	0.00040
2.411-2.471	Glutamine	0.00272	0.00021
2.471-2.501		0.00111	0.00008
2.501-2.551	Citrate	0.00111	0.00007
2.551-2.571	Methylamine	0.00060	0.00005
2.571-2.612		0.00170	0.00011
2.612-2.647		0.00285	0.00020
2.647-2.687	Citrate	0.00424	0.00031
2.647-2.705	Citrate	0.00641	0.00046
2.705-2.715		0.00133	0.00010
2.715-2.848	=CH-CH <sub>2</sub> -CH=	0.00849	0.00040
2.687-2.791	Citrate/=CH-CH <sub>2</sub> -CH=	0.00909	0.00063
2.791-2.831	Citrate/=CH-CH <sub>2</sub> -CH=	0.00164	0.00011
2.831-2.848	Citrate/=CH-CH <sub>2</sub> -CH=	0.00123	0.00010
2.848-2.887		0.00325	0.00026
2.848-2.873		0.00211	0.00017
2.873-2.927	Albumine	0.00521	0.00042
2.887-2.927		0.00408	0.00033
2.927-2.967		0.00548	0.00043
2.967-3.071	Creatine/albumin lysyl	0.01003	0.00077
2.967-2.98		0.00218	0.00017
2.98-3.028	Lysine	0.00539	0.00042
3.028-3.044	Creatine	0.00121	0.00010
3.044-3.071		0.00158	0.00012
3.071-3.111	Creatinine	0.00233	0.00013
3.111-3.129		0.00111	0.00007
3.129-3.167		0.01657	0.00141
3.167-3.269	Choline, glucose	0.02303	0.00174
3.269-3.311	Glucose	0.00337	0.00020
3.311-3.312	Glucose	0.00007	0.00001

3.312-3.367	Glucose	0.01065	0.00080
3.367-3.437	Glucose	0.01468	0.00112
3.437-3.496	Glucose	0.00896	0.00057
3.496-3.587	Glucose	0.02650	0.00102
3.587-3.616	Ethanol	0.06577	0.00304
3.616-3.711	Glutamine	0.03258	0.00145
3.711-3.713		0.00061	0.00005
3.713-3.791	Glucose	0.01459	0.00102
3.791-3.795	Glucose	0.00077	0.00006
3.795-3.86	Glucose	0.01559	0.00107
3.86-3.951	Creatine	0.00972	0.00067
3.86-3.919		0.00668	0.00047
3.919-3.932	Creatine	0.00147	0.00011
3.932-3.951		0.00179	0.00013
3.951-3.978		0.00177	0.00012
3.978-4.021		0.00330	0.00018
4.021-4.081	Creatine glyceryl	0.00871	0.00051
4.021-4.034		0.00149	0.00009
4.034-4.055	Creatinine	0.00337	0.00018
4.055-4.081	Choline	0.00408	0.00026
4.081-4.151	Lactate	0.00480	0.00037
4.151-4.184		0.00193	0.00013
4.184-4.198		0.00096	0.00005
4.198-4.208		0.00077	0.00004
4.208-4.311	Threonine	0.00702	0.00037
5.007-5.081		0.00454	0.00016
5.081-5.087		0.00039	0.00001
5.087-5.127		0.00227	0.00008
5.127-5.167		0.00260	0.00013
5.167-5.207	CH glyceryl	0.00726	0.00039
5.207-5.247	Glucose	0.00901	0.00080
5.247-5.351	Unsaturation	0.00959	0.00091
5.351-5.389	Unsaturation	0.00109	0.00004
5.389-5.431		0.00102	0.00004
5.431-5.471		0.00085	0.00003
5.471-5.51		0.00075	0.00003
5.51-5.528		0.00034	0.00001
5.528-5.63		0.00165	0.00006
5.63-5.634		0.00008	0.00000
5.634-5.647		0.00022	0.00001
5.647-5.687		0.00072	0.00005
5.687-5.727	Urea	0.00104	0.00011
5.727-5.831	Urea	0.00157	0.00011
5.831-5.864		0.00039	0.00002
5.864-5.889		0.00028	0.00001
5.889-5.969		0.00079	0.00003
5.969-6.07		0.00086	0.00004
6.07-6.1		0.00024	0.00001
6.1-6.133		0.00026	0.00001

6.133-6.17		0.00028	0.00001
6.17-6.207		0.00028	0.00001
6.207-6.248		0.00032	0.00002
6.248-6.34		0.00064	0.00003
6.34-6.367		0.00019	0.00001
6.367-6.407		0.00029	0.00002
6.407-6.447		0.00029	0.00002
6.447-6.492		0.00032	0.00002
6.492-6.527		0.00026	0.00002
6.527-6.567		0.00032	0.00002
6.567-6.613		0.00040	0.00003
6.613-6.648		0.00038	0.00003
6.648-6.687		0.00067	0.00006
6.687-6.728		0.00070	0.00006
6.728-6.767	Protein	0.00080	0.00007
6.767-6.857		0.00202	0.00015
6.857-6.943	Tyrosine	0.00171	0.00013
6.857-6.912	Tyrosine	0.00107	0.00009
6.912-6.943	Protein	0.00066	0.00005
6.943-6.968	Protein	0.00059	0.00005
6.968-7.023	1-methylhistidine	0.00163	0.00013
7.023-7.054	Histidine	0.00081	0.00006
7.054-7.071		0.00036	0.00003
6.968-7.071	Histidine	0.00276	0.00021
7.071-7.108		0.00076	0.00006
7.108-7.12	Protein	0.00026	0.00002
7.12-7.127		0.00024	0.00002
7.127-7.211	Tyrosine	0.00171	0.00013
7.128-7.159		0.00071	0.00006
7.159-7.211	Tyrosine	0.00099	0.00008
7.211-7.27		0.00106	0.00009
7.27-7.279		0.00018	0.00001
7.279-7.344	Phenylalanine	0.00110	0.00009
7.344-7.375		0.00056	0.00005
7.375-7.431	Phenylalanine	0.00061	0.00005
7.431-7.471		0.00033	0.00003
7.471-7.508		0.00031	0.00002
7.508-7.539	3-methylhistidine	0.00052	0.00003
7.539-7.629	3-methylhistidine	0.00123	0.00007
7.629-7.647	3-methylhistidine	0.00025	0.00002
7.647-7.687	1-methylhistidine	0.00073	0.00005
7.687-7.772	Histidine	0.00162	0.00010
7.772-7.831	1-methylhistidine	0.00066	0.00005
7.831-7.871		0.00025	0.00002
7.871-7.911		0.00023	0.00002
7.911-7.921		0.00006	0.00001
7.921-7.984		0.00036	0.00003
7.984-8.018		0.00015	0.00001
8.018-8.047		0.00012	0.00001

8.047-8.144	0.00051	0.00004
8.144-8.231	0.00036	0.00003
8.231-8.263	0.00010	0.00001
8.263-8.306	0.00013	0.00001
8.306-8.344	0.00011	0.00001
8.344-8.364	0.00006	0.00001
8.364-8.427	0.00021	0.00001
8.427-8.468	0.00009	0.00001
8.468-8.503	0.00008	0.00001
8.503-8.551	0.00010	0.00001

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**Supplementary Table 5.** Description of the 215 NOESY continuous variables used in statistical analyses, *Milieu Intérieur* study, France, 2012 (N=846).

Spectral region (ppm)	Metabolite assignment	Mean	SD
0.487-0.527		0.00383	0.00006
0.527-0.567		0.00451	0.00007
0.567-0.607		0.00581	0.00017
0.607-0.647	CH <sub>3</sub> cholesterol	0.00667	0.00019
0.647-0.687	CH <sub>3</sub> cholesterol	0.00724	0.00013
0.687-0.727	CH <sub>3</sub> cholesterol	0.00861	0.00013
0.727-0.767		0.01317	0.00037
0.767-0.807	CH <sub>3</sub> lipoproteins	0.02595	0.00239
0.807-0.911	CH <sub>3</sub> lipoproteins	0.04896	0.00336
0.911-0.95	Leucine valine	0.01381	0.00042
0.95-0.984	Leucine valine	0.01024	0.00032
0.984-1.021	Leucine valine	0.01050	0.00033
1.021-1.067	Leucine valine	0.01267	0.00052
0.97-1	Valine	0.00897	0.00026
1-1.021	Isoleucine	0.00592	0.00020
1.021-1.051	Valine	0.00830	0.00033
1.051-1.067		0.00464	0.00020
1.067-1.089		0.00647	0.00027
1.09-1.124		0.01154	0.00535
1.089-1.127		0.01296	0.00582
1.127-1.167	Isobutyrate	0.02197	0.00281
1.167-1.207	CH <sub>2</sub> lipoproteins	0.03355	0.00469
1.207-1.311	CH <sub>2</sub> lipoproteins	0.06990	0.01291
1.311-1.351	Lactate threonine	0.01167	0.00059
1.351-1.391	CH <sub>2</sub> lipoproteins	0.01139	0.00032
1.391-1.429	Lysine	0.01244	0.00037
1.429-1.447		0.00611	0.00020
1.447-1.522	Alanine	0.02360	0.00115
1.522-1.591	CH <sub>2</sub> CH <sub>2</sub> COOC	0.01775	0.00063
1.591-1.631	CH <sub>2</sub> CH <sub>2</sub> COOC	0.01008	0.00013
1.631-1.636	CH <sub>2</sub> CH <sub>2</sub> COOC	0.00156	0.00002
1.636-1.647	CH <sub>2</sub> CH <sub>2</sub> COOC	0.00317	0.00004
1.647-1.677		0.00818	0.00010
1.677-1.707	Arginine	0.00751	0.00009
1.707-1.751	Lysine	0.01026	0.00012
1.751-1.791		0.00921	0.00012
1.791-1.831		0.00906	0.00012
1.831-1.851		0.00479	0.00006
1.851-1.887		0.00888	0.00014
1.887-1.927	Acetate	0.01233	0.00048
1.927-1.967	Glycoproteins N-acetyl, CH <sub>2</sub> CH	0.01538	0.00116
1.927-2.018	CH <sub>2</sub> CH <sub>2</sub> COOC	0.03397	0.00208
2.018-2.057	Glycoproteins N-acetyl, CH <sub>2</sub> CH	0.00990	0.00020
2.057-2.151	Glutamine	0.02009	0.00043
2.064-2.11		0.01025	0.00016



2.11-2.151		0.00838	0.00029
2.151-2.174		0.00544	0.00045
2.174-2.264	Acetone, CH <sub>2</sub> CO	0.01879	0.00070
2.264-2.311		0.00825	0.00011
2.311-2.351		0.00623	0.00008
2.351-2.388	Pyruvate	0.00584	0.00011
2.351-2.374	Pyruvate	0.00357	0.00005
2.374-2.414		0.00640	0.00015
2.388-2.414		0.00414	0.00009
2.414-2.471	Glutamine	0.00704	0.00011
2.471-2.502		0	0.00337
2.502-2.551	Citrate	0.00454	0.00006
2.551-2.58	Methylamine	0.00272	0.00006
2.58-2.627		0.00502	0.00009
2.627-2.647		0.00263	0.00011
2.647-2.687	Citrate/=CH-CH <sub>2</sub> -CH=	0.00536	0.00034
2.687-2.791	Citrate/=CH-CH <sub>2</sub> -CH=	0.01113	0.00062
2.791-2.831	Citrate/=CH-CH <sub>2</sub> -CH=	0.00402	0.00007
2.831-2.84	Citrate/=CH-CH <sub>2</sub> -CH=	0.00113	0.00003
2.84-2.847	Citrate/=CH-CH <sub>2</sub> -CH=	0.00095	0.00003
2.647-2.708	Citrate	0.00811	0.00055
2.708-2.847	=CH-CH <sub>2</sub> -CH=	0.01415	0.00040
2.847-2.887		0.00510	0.00010
2.887-2.927		0.00573	0.00012
2.927-2.978		0.00821	0.00016
2.978-3.071	Creatine /albumin lysyl	0.01079	0.00020
3.071-3.111	Creatinine	0.00354	0.00005
3.111-3.13		0.00195	0.00004
3.13-3.167		0.00967	0.00102
3.167-3.27	Choline, glucose	0.01351	0.00070
3.27-3.311	Glucose	0.00306	0.00010
3.311-3.314	Glucose	0.00026	0.00001
3.314-3.366	Glucose	0.00686	0.00040
3.366-3.367	Glucose	0.00021	0.00001
3.367-3.438	Glucose	0.00941	0.00057
3.438-3.497	Glucose	0.00667	0.00027
3.497-3.532	Glucose	0.00421	0.00017
3.532-3.588	Glucose	0.00633	0.00107
3.588-3.611	Glucose	0.00403	0.00347
3.611-3.707		0.01605	0.00092
3.707-3.791	Glucose	0.01341	0.00049
3.791-3.793	Glucose	0.00046	0.00002
3.793-3.807	Glucose	0.00243	0.00008
3.807-3.86	Glucose	0.01081	0.00041
3.86-3.951	Creatine	0.01395	0.00025
3.861-3.918		0.00884	0.00019
3.918-3.95	Creatine	0.00497	0.00009
3.951-3.952		0.00028	0.00000
3.952-4.03		0.01085	0.00025

4.03-4.094	Creatinine glyceryl lactate	0.01032	0.00034
4.094-4.151	Creatinine glyceryl lactate	0.00693	0.00012
4.031-4.075	Creatinine glyceryl	0.00761	0.00031
4.075-4.151	Lactate	0.00953	0.00016
4.151-4.191		0.00458	0.00006
4.191-4.214		0.00282	0.00011
4.214-4.311	Threonine	0.00959	0.00018
5.031-5.042		0.00018	0.00001
5.042-5.047		0.00009	0.00001
5.047-5.087		0.00065	0.00004
5.087-5.127		0.00081	0.00006
5.127-5.167		0.00124	0.00015
5.167-5.207	CH glyceryl	0.00418	0.00038
5.207-5.247	Glucose	0.00576	0.00099
5.247-5.351	Unsaturation	0.00452	0.00098
5.351-5.391	Unsaturation	0.00052	0.00004
5.391-5.431		0.00042	0.00003
5.431-5.471		0.00037	0.00003
5.471-5.511		0.00035	0.00002
5.511-5.551		0.00034	0.00002
5.551-5.591		0.00032	0.00002
5.591-5.631		0.00033	0.00002
5.631-5.639		0.00008	0.00001
5.639-5.647		0.00008	0.00001
5.647-5.687		0.00039	0.00004
5.687-5.727		0.00051	0.00009
5.727-5.83	Urea	0.00095	0.00009
5.83-5.835		0.00006	0.00001
5.835-5.861		0.00026	0.00002
5.861-5.887		0.00026	0.00002
5.887-5.965		0.00071	0.00005
5.965-5.966		0.00002	0.00000
5.966-6.07		0.00102	0.00004
6.07-6.11		0.00038	0.00001
6.11-6.118		0.00009	0.00000
6.118-6.126		0.00009	0.00000
6.126-6.166		0.00042	0.00001
6.166-6.208		0.00047	0.00002
6.208-6.246		0.00054	0.00002
6.246-6.286		0.00060	0.00002
6.286-6.327		0.00066	0.00002
6.327-6.366		0.00073	0.00002
6.366-6.406		0.00091	0.00002
6.406-6.446		0.00102	0.00002
6.446-6.486		0.00103	0.00002
6.486-6.526		0.00114	0.00003
6.526-6.566		0.00129	0.00003
6.566-6.606		0.00137	0.00004
6.606-6.646		0.00150	0.00005

6.646-6.686		0.00191	0.00006
6.686-6.726		0.00213	0.00006
6.726-6.766	Protein	0.00222	0.00007
6.766-6.863		0.00554	0.00014
6.863-6.905	Tyrosine	0.00258	0.00007
6.905-6.926		0.00138	0.00004
6.926-6.966		0.00274	0.00007
6.966-7.007		0.00315	0.00009
7.007-7.11	Histidine	0.00701	0.00019
7.11-7.15		0.00254	0.00005
7.15-7.183	Tyrosine	0.00207	0.00004
7.183-7.209	Tyrosine	0.00157	0.00004
7.209-7.27		0.00350	0.00007
7.27-7.276		0.00037	0.00001
7.276-7.35	Phenylalanine	0.00369	0.00009
7.35-7.367		0.00087	0.00002
7.367-7.43	Phenylalanine	0.00264	0.00008
7.43-7.47		0.00152	0.00006
7.47-7.51		0.00147	0.00005
7.51-7.543		0.00126	0.00004
7.543-7.566	3-methylhistidine	0.00094	0.00003
7.566-7.606	3-methylhistidine	0.00162	0.00006
7.606-7.646	3-methylhistidine	0.00168	0.00005
7.646-7.686	1-methylhistidine	0.00168	0.00005
7.686-7.772	Histidine	0.00366	0.00011
7.772-7.83	1-methylhistidine	0.00214	0.00006
7.83-7.87		0.00138	0.00004
7.87-7.908		0.00134	0.00004
7.908-7.917		0.00035	0.00001
7.917-7.926		0.00035	0.00001
7.926-7.966		0.00150	0.00007
7.966-8.006		0.00155	0.00007
8.006-8.046		0.00161	0.00006
8.046-8.086		0.00165	0.00006
8.086-8.157		0.00293	0.00010
8.157-8.23		0.00289	0.00010
8.23-8.27		0.00151	0.00005
8.27-8.31		0.00145	0.00005
8.31-8.35		0.00140	0.00004
8.35-8.39		0.00135	0.00004
8.39-8.431		0.00133	0.00004
8.431-8.471	Formic acid	0.00120	0.00003
8.471-8.511		0.00120	0.00003
8.511-8.551		0.00108	0.00003
8.551-8.591		0.00105	0.00003
8.591-8.631		0.00095	0.00003
8.631-8.671		0.00099	0.00003
8.671-8.71		0.00093	0.00003
8.71-8.751		0.00094	0.00004

8.751-8.791	0.00088	0.00004
8.791-8.831	0.00084	0.00004
8.831-8.871	0.00079	0.00003
8.871-8.911	0.00073	0.00003
8.911-8.951	0.00067	0.00003
8.951-8.991	0.00060	0.00002
8.991-9.031	0.00054	0.00002
9.031-9.071	0.00049	0.00002
9.071-9.111	0.00045	0.00002
9.111-9.151	0.00041	0.00002
9.151-9.191	0.00038	0.00002
9.191-9.231	0.00036	0.00002
9.231-9.271	0.00034	0.00002
9.271-9.311	0.00031	0.00002
9.311-9.351	0.00028	0.00002
9.351-9.391	0.00026	0.00001
9.391-9.431	0.00024	0.00001
9.431-9.471	0.00022	0.00001
9.471-9.511	0.00021	0.00001

**Supplementary Table 6.** Associations between covariates and metabolomic spectra (CPMG and NOESY sequences), using Principal Component Partial R-square (PC-PR2), *Milieu Intérieur* study, France, 2012 (N=846).

Covariate	CPMG sequence		NOESY sequence	
	R <sup>2</sup>	p-value <sup>1</sup>	R <sup>2</sup>	p-value <sup>1</sup>
Age	0.02	<0.0001	0.05	<0.0001
Sex	0.05	<0.0001	0.03	<0.0001
BMI	0.03	<0.0001	0.02	<0.0001
Smoking status	0.01	0.5	0.03	0.02
Physical activity	0.002	0.8	0.001	0.4

<sup>1</sup> p-value for PC-PR2 analysis (Fages A, et al. Batch profiling calibration for robust NMR metabonomic data analysis. *Anal. Bioanal. Chem.* 405 (27): 8819-8827).

**Supplementary Table 7.** Associations between NMR variables and  $\alpha$ -diversity indexes with a Q-value  $\leq 0.1$  after multiple-testing correction, using Spearman partial correlations, *Milieu Intérieur* study, France, 2012 (N=846).

Sequence	Shift (ppm)	Assignment	Observed richness			Chao1 richness estimate			
			$\rho$	p-value <sup>1</sup>	Q-value <sup>2</sup>	$\rho$	p-value <sup>1</sup>	Q-value <sup>2</sup>	
CPMG	0.567-0.607	Unassigned signal	0.090	0.009	0.09	0.083	0.02	0.12	
	1.207-1.311	Lipoproteins, -CH <sub>2</sub>	-0.105	0.002	0.05	-0.103	0.003	0.08	
	1.521-1.527	CH <sub>2</sub> CH <sub>2</sub> COOC	-0.101	0.003	0.05	-0.097	0.005	0.08	
	2.151-2.166	Lipid, -CH <sub>2</sub> =C=C-	-0.097	0.005	0.06	-0.099	0.004	0.08	
	2.166-2.167		-0.101	0.003	0.05	-0.103	0.003	0.08	
	2.167-2.264	Acetone, CH <sub>2</sub> CO	-0.108	0.002	0.05	-0.092	0.008	0.09	
	2.551-2.571	Methylamine	0.118	0.001	0.04	0.105	0.002	0.08	
	2.831-2.848	Citrate/=CH-CH <sub>2</sub> -CH=	0.088	0.01	0.09	0.077	0.03	0.12	
	3.071-3.111	Creatinine	0.109	0.002	0.05	0.095	0.006	0.08	
	3.167-3.269	Choline and glucose	0.095	0.006	0.06	0.073	0.03	0.14	
	3.311-3.312	Glucose	0.095	0.006	0.06	0.081	0.02	0.12	
	3.616-3.711	Glutamine	0.087	0.012	0.1	0.094	0.007	0.08	
	5.127-5.167	Unassigned signal	-0.105	0.002	0.05	-0.104	0.003	0.08	
	5.864-5.889		0.088	0.01	0.09	0.077	0.03	0.12	
	5.889-5.969		0.100	0.004	0.05	0.079	0.02	0.12	
	6.492-6.527		0.099	0.004	0.05	0.091	0.008	0.09	
	6.728-6.767	Proteins	0.108	0.002	0.05	0.100	0.004	0.08	
	6.857-6.943	Tyrosine	0.093	0.007	0.07	0.091	0.008	0.09	
	6.912-6.943	Proteins	0.103	0.003	0.05	0.095	0.006	0.08	
	6.943-6.968		0.121	0.0005	0.04	0.109	0.002	0.08	
7.508-7.539	3-methylhistidine	0.099	0.004	0.05	0.084	0.01	0.12		
7.629-7.647		0.102	0.003	0.05	0.095	0.006	0.08		
7.647-7.687	1-methylhistidine	0.126	0.0002	0.04	0.105	0.002	0.08		
7.687-7.772	Histidine	0.110	0.001	0.05	0.107	0.002	0.08		
NOESY	4.191-4.214	Unassigned signal	-0.133	0.0001	0.02	-0.118	0.0006	0.04	
	5.830-5.835		-0.112	0.001	0.05	-0.100	0.004	0.1	
	5.861-5.887		0.114	0.001	0.05	0.109	0.002	0.05	
	5.887-5.965		0.126	0.0003	0.03	0.117	0.0007	0.04	
	5.965-5.966		0.110	0.001	0.05	0.117	0.0007	0.04	
	6.486-6.526		0.104	0.003	0.08	0.096	0.005	0.12	
	6.726-6.766		Proteins	0.122	0.0004	0.03	0.119	0.001	0.04
	7.646-7.686		1-methylhistidine	0.101	0.003	0.09	0.086	0.01	0.18

<sup>1</sup> p-value for non-parametric Spearman partial correlations (adjusted for age, sex, BMI, smoking status, physical activity, and sequencing depth) before Benjamini-Hochberg correction.

<sup>2</sup> Corrected p-value (Benjamini-Hochberg method, 10%-FDR). Only associations with Q-value  $\leq 0.1$  for observed richness, as well as subsequent associations with Chao1 richness are presented.