

1 **Association of subclinical mastitis prevalence**

2 **with sheep breeds reared in Greece**

3
4 N.G.C. Vasileiou¹, D.A. Gougoulis¹, V. Riggio², K.S. Ioannidi¹,
5 D.C. Chatzopoulos¹, V.S. Mavrogianni², E. Petinaki³, G.C. Fthenakis^{1*}

6
7
8
9 **Supplementary material 1.** Detailed description of procedures and techniques employed in the
10 study.

11 1. *Sheep farms and animal sampling*

12 In total, 111 sheep farms in the 13 administrative regions of Greece were included into the
13 study and visited for collection of samples and information. Veterinarians active in small ruminant
14 health management around Greece, were contacted and asked if they wished to collaborate in the
15 investigation. In total, 23 veterinarians had agreed to collaborate. Farms were selected by the
16 collaborating veterinarians on convenience basis (i.e., willingness of farmers to accept a visit by
17 University personnel for sampling animals). The principal investigators (NGCV, GCF) visited all
18 farms for sample collection. Farms were classified according to management system followed
19 therein, as intensive (n=26), semi-intensive (n=57), semi-extensive or extensive (n=28), by following
20 the criteria of the European Food Safety Authority (2014).

21 In each farm, 20 clinically healthy ewes (*secundiparae* or older) were selected at random for
22 sampling. For selection of animals, farmers had been asked to remove *primiparae* ewes and ewes
23 with known udder abnormalities from the main flock. A standardised clinical examination
24 (observation, palpation, comparison between glands) of the udder was performed, always by the
25 principal investigator (NGCV) (Fthenakis, 1994; Mavrogianni et al., 2005) and the first two squirts
26 of secretion were drawn on the gloved hand of an assisting investigator and assessed. All
27 investigators involved in sampling procedures wore disposable, non-sterile latex gloves. If udder
28 abnormalities were recorded during clinical examination, the ewe was excluded from sampling.
29 Animals found with abnormalities and excluded, were not replaced.

30 Standard methods for aseptic collection of milk samples were followed (Fthenakis, 1994).
31 Then, 10 to 15 mL of secretion were collected into a sterile container; separate samples were
32 collected from each mammary gland into separate containers. Milk samples were then drawn onto
33 a paddle for performing the California Mastitis Test (CMT). For transportation, samples were
34 stored into portable refrigerators with ice packs and transported by car; for samples collected in
35 islands, airplane or boat transportation, as accompanying luggage, was also involved.

36 2. *Paraclinical examinations*

37 Laboratory procedures started within 24 h after collection. Milk samples (10 µL) were
38 cultured using Columbia blood agar plates incubated aerobically at 37 °C for up to 72 h. Bacterial

39 identifications were performed by using standards methods (Barrow and Feltham, 1993; Euzeby,
40 1997).

41 After sample collection, at ewe-side, all samples were tested by use of the CMT. The test was
42 performed as previously described for ewes' milk (Fthenakis, 1995); it was carried out and scored
43 always by the same person, i.e., the principal investigator (NGCV). Five degrees of reaction
44 ('negative', 'trace', '1', '2', '3') were described (Schalm et al., 1971). Milk smears were also produced
45 and dried. The milk smears were stained by the Giemsa method for estimation of leucocyte
46 subpopulations; proportion of leucocyte types therein was calculated by observing at least 10
47 fields of each milk film under magnification 10 \times . Subsequently, the Microscopic cell counting
48 method (Mccm) (IDF reference method) (International Dairy Federation, 1984; Contreras et al.,
49 2007; Raynal-Ljutovac et al., 2007) was performed in 894 samples (20.3% of all samples).

50 3. *Data management and analysis*

51 Ewes were considered to have subclinical mastitis when a bacteriologically positive milk
52 sample ([a] >10 colonies of the same organism and [b] no more than two different types of colonies)
53 with concurrently increased CMT score (≥ 1) plus neutrophil and lymphocyte proportion ($\geq 65\%$ of
54 all leucocytes) was detected (Fragkou et al., 2014). The definition referred to ewes (hence, animals
55 with both glands affected were counted as one case).

56 Quantitative information on the cellular content of ewes' milk was obtained by using two
57 sets of data: the CMT results and the results of the Mccm. Although it is generally established
58 that CMT results are reliable proxy measurements for somatic cell counts (SCCs) (Fthenakis,
59 1995; Gonzalez-Rodriguez and Carmenes, 1996), we further confirmed that in the present study.
60 Following assignment of numerical values to CMT scores (value 0 to score 'negative', value 1 to
61 score 'trace', value 2 to score '1', value 3 to score '2', and value 4 to score '3') and log₁₀-
62 transformations, correlation between CMT scores and Mccm SCCs was $r=0.913$ (95% CI: 0.902-
63 0.923) ($P<0.001$) and the corrected R^2 was 83.4%; significance of the difference between r and ρ
64 (the correlation hypothesized to exist within the population from which the sample had been
65 drawn) was $P<0.001$.

66 For analysis, data were entered into Microsoft Excel and analysed using IBM SPSS Statistics
67 (ver. 21) (IBM; Armonk, NY, USA). The outcome of 'subclinical mastitis' was considered. Exact
68 binomial confidence intervals (C.I.) were obtained. A preliminary assessment of the importance of
69 predictors was performed using by cross-tabulation with the chi-square test, and with simple
70 logistic regression without random effects. Subsequently, mixed-effects logistic regression was
71 employed to perform the same comparisons, using the different farms ($n=111$) as a 'random effect'.
72 Then, analysis of variance was employed and the following comparisons were made between farms
73 in relation to this outcome:

- 74 (a) farms with pure-bred animals *versus* farms with cross-bred animals,
- 75 (b) farms with Greek pure-bred animals *versus* farms with imported pure-bred animals,
- 76 (c) farms with imported pure-bred animals *versus* all other farms (i.e., farms with Greek pure-bred
77 animals and farms with cross-bred animals),

78 (d) farms with the various Greek pure-bred animals (in total, 8 breeds), farms with imported pure-
79 bred animals (in total, 2 breeds) and farms with cross-bred animals and
80 (e) farms with the various pure-bred animals (in total, 10 breeds) between them.

81 Subsequently, farms with the Greek breeds Cephalonia, Crete, Karagouniko, Karystos,
82 Lesvos and Vlahiko were considered together in a cluster termed 'Greek traditional indigenous
83 breeds' (n=18 farms), as initial comparison between those farms did not show significant
84 difference. Then, comparisons between the various breeds were repeated with smaller number of
85 breeds (in total, 3 Greek pure-breeds and 5 breeds in total).

86 Finally, a multivariable model was created using mixed-effects logistic regression with farm
87 as the random effect, which included as variables the management system in farms and the sheep
88 breed. The analysis was repeated by considering farms under semi-extensive and extensive
89 management clustered together (i.e., using 3 categories in the management system).

90 Statistical significance was defined at ≤ 0.05 .

91 4. References

- 92 Barrow GI, Feltham RKA 1993 *Manual for the Identification of Medical Bacteria*, 3rd edn. Cambridge
93 University Press. Cambridge, United Kingdom.
- 94 Contreras A, Sierra D, Sanchez A, Corrales JC, Marco JC, Paape MJ, Gonzalo C 2007 Mastitis in small
95 ruminants. *Small Ruminant Research* **68** 145-153.
- 96 European Food Safety Authority 2014 Scientific opinion on the welfare risks related to the farming of sheep
97 for wool, meat and milk production. *EFSA Journal* **12** 3933-4060.
- 98 Euzeby JP 1997 List of bacterial names with standing in nomenclature: a folder available on the Internet.
99 *International Journal of Systematic Bacteriology* **47** 590-592.
- 100 Fragkou IA, Boscós CM, Fthenakis GC 2014 Diagnosis of clinical or subclinical mastitis in ewes. *Small*
101 *Ruminant Research* **118** 86-92.
- 102 Fthenakis GC 1994 Prevalence and aetiology of subclinical mastitis in ewes of southern Greece. *Small*
103 *Ruminant Research* **13** 293-300.
- 104 Fthenakis GC 1995 California Mastitis Test and Whiteside Test in diagnosis of subclinical mastitis of dairy
105 ewes. *Small Ruminant Research* **16** 271-276.
- 106 Gonzalez-Rodríguez MC, Carmenes P 1996 Evaluation of the California mastitis test as a discriminant
107 method to detect subclinical mastitis in ewes. *Small Ruminant Research* **21** 245-250.
- 108 International Dairy Federation 1984 Recommended methods for somatic cell counting in milk. *Bulletin of the*
109 *International Dairy Federation* **168**.
- 110 Mavrogianni VS, Fthenakis GC, Brooks H, Papaioannou N, Cripps PJ, Taitzoglou I, Brellou G, Saratsis P
111 2005 The effects of inoculation of *Mannheimia haemolytica* into the teat of lactating ewes. *Veterinary*
112 *Research* **36** 13-25.
- 113 Raynal-Ljutovac K, Pirisi A, de Cremoux R, Gonzalo C 2007 Somatic cells of goat and sheep milk: analytical,
114 sanitary, productive and technological aspects. *Small Ruminant Research* **68** 126-144.
- 115 Schalm OW, Carroll EJ, Jain NC 1971 *Bovine Mastitis*. Lea and Febiger, Philadelphia, USA.

116
117
118
119



121
122
123
124
125
126
127
128

129 **Supplementary material 3.** Breeds in sheep farms in Greece according to management system
 130 applied in farms.

Sheep breeds	Management system (no. of farms)			Total
	Intensive	Semi-intensive	Semi-extensive or extensive	
1. Pure-breeds	17	25	16	58
1.1. Greek breeds	6	13	14	33
1.1.1. Cephalonia		1	1	2
1.1.2. Chios	6	4	3	13
1.1.3. Crete			4	4
1.1.4. Frisarta		2		2
1.1.5. Karagouniko		2	1	3
1.1.6. Karystos			1	1
1.1.7. Lesvos		4	1	5
1.1.8. Vlahiko			3	3
1.2. Imported breeds	11	12	2	25
1.2.1. Assaf	1	1		2
1.2.2. Lacaune	10	11	2	23
2. Cross-breeds	9	32	12	53
Total	26	57	28	111

131
 132
 133
 134
 135