

## 1 Detailed Materials & methods

### 2 *Animal and milk production records*

3 Mixed milk (volume in morning: noon: night = 4:3:3) samples were collected  
4 monthly from June 2010 to December 2014. Milk samples were treated with  
5 potassium bichromate (30 mg/tube) immediately after milking, and analyzed for SCC  
6 based on flow cytometry (Fossmatic 5000, Foss Electric, Denmark). The infrared  
7 technique (Milkoscan 6000, Foss Electric, Denmark) was used to determine  
8 concentrations of fat, protein, lactose, total solids (**TS**) and milk urea nitrogen (**MUN**).  
9 Production traits included test-day milk yield (**TDMY**), fat content (**FC**), protein  
10 content (**PC**), lactose content (**LC**), MUN and somatic cell count (**SCC**). Somatic cell  
11 score (**SCS**) was calculated using the formula:  $SCS = \log_2^{(SCC/100\ 000)} + 3$  (Wiggans et al.  
12 1987). These records were obtained from the Dairy Herd Improvement (**DHI**) lab of  
13 the Shanghai Dairy Cattle Breeding Center. Editing of the data was performed to  
14 ensure both reliability and consistency for statistical analyses. Requirements were  
15 designed as follows: TDMY between 5-60 kg, FC between 2-7%, PC between 2-6%,  
16 LC between 2-5.5%, TS content between 9-18%, MUN between 5-30%, SCC  
17 between  $1 \times 10^3$  -  $9999 \times 10^3$ , and SCS between 0 - 9. Only records from parities from 1  
18 to 3, and from 5 to 305 days in milk (DIM) were included. For analyses, 20 556  
19 records were retained and utilized in this study. The mean values and standard errors  
20 for the analyzed traits stratified by parity are summarized in Table 1.

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### 22 *DNA extraction*

23 Blood samples were placed immediately on ice for subsequent DNA extraction. An  
24 Eppendorf Biophotometer (Berlin, Germany) was used to assess the DNA  
25 concentration and DNA quality based on absorbance of ultraviolet light at 260 and  
26 280 nm. The ratio of  $OD_{260}/OD_{280}$  for the DNA samples ranged from 1.70 to 1.85, and  
27 the concentration of the DNA was above  $50\text{ ng }\mu\text{l}^{-1}$ . Finally, the concentration of the  
28 DNA sample used for PCR was diluted to  $50\text{ ng }\mu\text{l}^{-1}$  with ddH<sub>2</sub>O and stored at -20

29 °C.

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### 31 *SNP genotyping and transcription-factor binding sites analysis*

32 Primers for PCR and single base extension for the SNPs of *TLR4* c.-226 G>C and  
33 *TLR4* c.2021 C>T were designed using the Assay Designer software package  
34 (Sequenom Inc., San Diego, CA) (Table 2). As a quality control measure, genomic  
35 DNA from 40 animals was genotyped in duplicate for each SNP. Concordance across  
36 SNP and all duplicates was 100%.

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### 38 *SNP and Haplotype associations*

39 The additive (a) and dominance (d) and effects of single SNPs were evaluated using  
40 the equations:  $a = (AA-BB)/2$ , and  $d = AB-(AA+ BB)/2$ , where AA and BB indicate  
41 the two homozygous genotypes, AB represents the heterozygous genotype (Falconer  
42 and Mackay 1996). Allele substitution effects ( $\alpha$ ) at each SNP-locus were estimated  
43 by regressing the milk production traits and SCS on the number of copies of one allele  
44 of a SNP (Sherman et al., 2008) using a linear model in SAS. For the analyses above,  
45 the Bonferroni method was adopted to correct for multiple t-tests according to the  
46 number of SNP loci or haplotype blocks detected. We declared a significant SNP or  
47 haplotype if a raw *P*-value is  $<0.05/N$ , where N is the number of SNP loci or  
48 haplotype blocks tested in the analyses.

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59 **Applied tables**

60 **Table 1** Descriptive statistics for milking traits by parity (means ± SE)

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Parity	Records number	TDMY <sup>a</sup> (kg)	FC <sup>a</sup> (%)	PC <sup>a</sup> (%)	FPR <sup>a</sup>	SCS <sup>a</sup>	LC <sup>a</sup> (%)	TS <sup>a</sup> (%)	MUN <sup>a</sup> (g/100mL)
1	7 018	29 06±0 08	4 14±0 01	3 34±0 01	1 25±0 01	2 03±0 02	5 01±0 01	13 49±0 01	10 94±0 03
2	6 741	33 06±0 12	4 36±0 01	3 38±0 01	1 29±0 01	1 82±0 02	4 94±0 01	14 05±0 02	12 52±0 04
3	6 797	34 62±0 16	4 24±0 01	3 32±0 01	1 29±0 01	2 22±0 02	4 86±0 01	13 80±0 02	13 80±0 04
Total	20 556	32 21±0 07	4 25±0 01	3 35±0 01	1 27±0 01	2 02±0 01	4 95±0 01	13 77±0 01	12 17±0 02

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63 **Abbreviations:**

64 TDMY: test-day milk yield

65 FC: fat content

66 PC: protein content

67 FPR: fat to protein ratio

68 SCS: somatic cell score

69 LC: lactose content

70 TS: total solid

71 MUN: milk urea nitrogen

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74 **Table 2** The primers sequence for TLR4 c -226 G>C and c 2021 C>T

Loci	Primer of sequences(5'→3')	75
c -226 G>C	F:ACGTTGGATGGGTCTGCAGACGTTTTCTTC	76
	R:ACGTTGGATGTCTGGACTTTCGTTTCTCTG	77
	U: ATCCTCTAACTTCCCCTC	78
c 2021 C>T	F:ACGTTGGATGCTCGAGTAGATGACAAAGGC	79
	R:ACGTTGGATGTTCCACCTGATGCTTCTTGC	80
	U: GATGACAAAGGCATCATAG	81

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85 **Table 3** Genotypic and allelic frequency & values of  $X^2$  test significance for *TLR4* c

86 -226 G&gt;C and c 2021 C&gt;T in Holstein cows

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Locus	Genotype	Genotypic frequency	Number	Allele	Allelic frequency	$X^2$ -test (P value)
c -226 G>C (rs 29017188)	CC	0 199	172	C	0 447	0 017*
	CG	0 497	430	G	0 553	
	GG	0 305	264			
c 2021 C>T (rs 8193069)	CC	0 753	652	C	0 865	0 940
	CT	0 225	195	T	0 135	
	TT	0 022	19			

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89 \*:  $P < 0.05$ 

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91 **Table 4** Haplotype reconstruction for SNP in *TLR4* c -226 G>C and *TLR4* c 2021

92 C&gt;T and their frequencies

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Haplotypes	TLR4 c -226	TLR4 c 2021	Number	Frequencies
1	C	C	541	0 312
2	C	T	233	0 135
3	G	C	958	0 553
Total			1 732	1 000

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