

1 Table 1. Overview of rodent studies, clinical trials and *in vitro* assays addressing BAT-mediated thermogenesis by phytochemicals and their impact
 2 on energy expenditure and weight control.
 3

<i>Pterostilbene</i>	blueberries, berries, wine, wine grapes, sorghum ⁽¹⁾			
Reference and model	Design	Sex and N	Treatment	Outcome
(2) Golden Syrian Hamsters	DI	male n=8-10 per group	2.5 mg/kg BW/d in HFD 3 weeks	-BW plasma ↓C, ↓LDL, ↓LDL:HDL
(2) rat hepatic H4IIEC3 cells		male	1,10,100 or 300 μM PTS 30 min	↑PPARα reporter activity
(3) Zucker fa/fa rats	DI	male n=10 per group	15 mg/kg BW/d in HFD 6 weeks	↓BW, ↓fat mass plasma ↓C, ↓insulin
(4) Wistar rats	DI	male n=9 per group	15 or 30 mg/kg BW/d in HFHS 6 weeks	↓BW gain, ↓total fat mass, ↓vWAT, ↓sWAT ↑hepatic CPT1α and ACOX activity
(5) Zucker fa/fa rats	orogastric catheter	male n=10 per group	0, 15 or 30 mg/kg BW/d in chow diet, 6 weeks	↓BW, ↓AT weight ↑iBAT mRNA/protein: NRF1, UCP1, PPARα ↑iBAT CPT1b activity
(6) OLETF rats	DI	male n=6 per group	300 mg/kg BW/d in chow 4 weeks,	-BW, ↓abdominal WAT, ↓total WAT ↓RER, ↑FO, ↑EE ↓WAT FAS mRNA
(7) C57BL/6 mice	DI	male and female n=8 per group/sex	90 mg/kg BW/d in HFD 30 weeks	↓BW, ↑glucose tolerance (after 18 weeks) ↑iWAT thermogenic genes (PPARγ, PGC1α, SIRT1, CIDEA, TBX1), ↑iWAT UCP1 protein ➤ effects in m+f, stronger than in f vs m
(7) 3T3-L1 adipocytes	acute	male	5 μM PTS for 24 hours, d12	↑UCP1 protein, ↑CIDEA, FGF21 mRNA

(8) hypercholesterolaemic Caucasian, AA	placebo parallel	male and female 100 mg/d, n=20 (m5/f15) 250 mg/d, n=20 (m6/f14) placebo, n=20 (m7/f13)	100 or 250 mg/d 6-8 weeks	-BMI (if stratified for C medication ↓BMI) plasma ↑total C, ↑LDL, -HDL
Resveratrol	peanuts and peanut products, grapes, red wine, soy, herbal remedies ⁽⁹⁾			
(10) C57Bl/6J mice	DI	male n=8-10 per group	400 mg/kg BW/d in HFD 15 weeks	↓weight gain, ↓final BW, ↓WAT mass ↑VO ₂ , ↑cold-resistance (rectal T) ↑iBAT mito content, ↑iBAT thermogenic genes (UCP1, PGC1 α , PPAR α) ↑mito gene enrichment in muscle, ↑IS
(11) C57BL/6NIA	DI	male n=6-9 per group	22.4 mg/kg BW/d in HFD middle aged mice, for 55 weeks	-fat distribution, -BW (trend for↓), ↑IS, -BT ↑survival plasma -TAG, ↑C, ↓GLC, ↓insulin
(12) grey mouse lemur (<i>Microcebus murinus</i>)	DI Before-after	male n=6	200 mg/kg BW/d 4 weeks	↑RER, ↓weight gain vs baseline week ↓food intake, -locomotor activity
(13) mice (strain unknown)	DI	male n=8 per group	0.4% w/w in chow diet 8 weeks	↓WAT mass, -BW, ↑VO ₂ , -locomotor activity ↑iBAT thermogenic genes (UCP1, PRDM16, SIRT1), plasma -C, -TAG, -GLC
(14) CD-1 mice	DI	female n=6 per group	0.1% w/w in HFD 4 weeks	↓BW gain, ↑VO ₂ , ↓RER, ↑EE (p=0.065), ↑UCP1+ in iWAT sections, ↓iWAT adipocyte size ↑pAMPK, UCP1, PRMD16 protein in iWAT plasma ↓insulin, ↓TAG, -GLC

(14) SVF from iWAT of CD-1 mice	long-term	unknown	10 μ M RSV during differentiation with brown adipogenic cocktail	↑thermogenic genes (UCP1, ELOVL3, PGC1 α , CIDEA, PRDM16), ↑UCP1, PRDM16 protein ↑respiration, ↑pAMPK ➤ effects AMPK-dependent
(15) CD-1 mice	DI	female	0.1% RSV w/w in HFD 4 weeks	↓BW gain, ↑BA number in iBAT sections ↑iBAT UCP1, PRDM16 and pAMPK protein
(16) Spargue-Dawley rats	DI	male n=8 per group	30 mg/kg BW/d in HFHS 6 weeks	-BW, ↓fat mass ↑BAT thermogenic genes (UCP1, PGC1 α , TFAM) ↑iBAT UCP1 protein, ↓acetylated PGC1 α (muscle)
(17) obese, healthy (BMI>30) Denmark	placebo parallel	male n=12 per group	500 mg RSV or placebo tablet/d 4 weeks	-BW, -total fat mass, -vWAT mass, plasma -GLC, -C, -TAG, -ALT, -HbA1c -acetylated lysine, pAMPK in muscle
(18) non-obese, normoglycaemic Caucasian	placebo parallel	female RSV n=15 placebo n=14	75 mg RSV/d 12 weeks	-BW, -fat mass, -sWAT mass, -vWAT mass plasma -leptin, -C, -TAG, -NEFA, -GLC -REE, -BP, -IS -WAT microarray: -mito function, -FO genes -muscle SIRT1 activity
(19) older adults with IGT US	before-after	male n=3 female n=7	1, 1.5 or 2 g RSV/d 4 weeks	-BW, -fat mass, -BP plasma -C, -TAG, -insulin, -ALT, -insulin ↓post-meal GLC, ↓post-meal insulin
(20) obese, healthy men	placebo crossover	male n=11	150 mg RSV/d (resVida) or placebo, 30 days per treatment	-BW, -fat mass ↓SEE (caloric restriction), ↑diurnal RER muscle ↑pAMPK, ↑mito activity, ↑SIRT1, PGC1 α protein
Quercetin	apples, onions, black currants, red wine, black tea, nuts, seeds, shallots ⁽²¹⁾			
(22)	DI	male	0.36% or 0.72% w/w OPE in	↓BW, ↓intra-abdominal fat mass

Sprague–Dawley rats		n=7 per group	HFD, 8 weeks	
(23) Wistar rats	DI	male n=12 per group	185, 270, 925 mg/kg BW/d in HFD 8 weeks	↓BW gain, ↓total fat mass, ↓vWAT plasma ↓TAG, ↓NEFA, -C, ↓GLC at high dose ↓hepatic fat content, ↑fecal lipids ↑PPAR α , ↑SIRT1, ↓ACC, ↓FAS mRNA (WAT)
(24) Zucker fa/fa rats	oral gavage	male n=7 per group	10 mg/kg BW/d RSV or vehicle with HFD 10 weeks	↓BW gain plasma ↓TAG, ↓C, ↓GLC, ↓insulin, ↓HOMA-IR ↓TNF α production, iNOs protein in vWAT
(25) C57BL/6	DI	male n=6 for OPE n=9 for control	0.5% w/w OPE in HFD 8 weeks	-BW gain, -eWAT mass, -rWAT mass ↑thermogenic genes (UCP1, PRDM16, CIDEA, PGC1 α) in rWAT
(25) 3T3-L1 adipocytes	long-term	male	100 μ M quercetin at d5, 7, 9	↑UCP1, SIRT1, PGC1 α mRNA and protein ↑pAMPK, pHSL, ↓lipogenic genes (FAS, ACC)
(26) C57BL/6	DI	male n=8-10 per group	0.1% w/w quercetin in HFD 8 weeks	-BW, -fat mass, -EE, -RER, -FO, -CHO, ↓plasma TAG ↑sWAT thermogenic genes (UCP1, ELOVL3) ↑UCP1+ cells in sWAT, ↑FA uptake in sWAT -mito content sWAT, -BAT morphology/genes
(27) C57BL/6J	DI	male n=8 per group	0.8% w/w in HFD 8 weeks	-BW, -adiposity, -EE, -RER ↓plasma inflammatory cytokines (INF γ , IL1, IL4)
(28) C57BL/6	DI	male n=6 per group	0.05% w/w quercetin in HFD 9 weeks	-BW, ↓WAT adipocyte size ↑UCP1+ cells, ↑UCP1, ↑PGC1 α protein in iWAT ↑iWAT thermogenic genes (UCP1, PRDM16, MEM26, NRF-1) ↑PKA, pAMPK protein in iWAT ↑plasma NE, ↑iBAT UCP1

(29) isolated Wistar rat adipocytes	acute	male	0, 1, 10, 100 or 250 μ M quercetin, 15 min	\uparrow PDE activity, \uparrow cAMP, \uparrow epinephrine-stimulated lipolysis
(30) C57BL/6	DI	male n=8 per group	0.1% w/w quercetin in HFD 12-17 weeks	\downarrow BW, \downarrow sWAT mass, \downarrow eWAT mass, \downarrow eWAT cell size, plasma \downarrow leptin, \downarrow insulin, \downarrow TNF α , \downarrow IL6 \downarrow mast cell infiltration eWAT \downarrow eWAT TNF α , IL6 \uparrow eWAT SIRT1, pAMPK protein \uparrow iBAT UCP1 mRNA,
(31) university students healthy Korean	placebo crossover	female n=12	100 mg/d quercetin or placebo capsule for 2 weeks each	-BW, -fat mass, -WHR, -BMI, -SBP, -DBP plasma -TAG, -C, -LDL
(32) overweight/obese subjects (BMI>23 kg/m ²) Korean	placebo crossover	female quercetin n=18 placebo n=19	100 mg/d quercetin or placebo capsules 12 weeks	-BW, -fat mass, -BMI plasma -ALT,-leptin, \uparrow adiponectin, -TNF α , -IL4
(33) overweight/obese subjects Korean	placebo crossover	male n=5 female n=31	100 mg/d quercetin or placebo capsules 12 weeks	-BW, -fat mas, -BMI, -WC, -RER, -REE plasma -GLC, -C, -LDL, \downarrow TAG, -leptin -before-after effects on REE, BW, RER, BMI within quercetin group
(34) Meta-analysis	9 RCTs	male=189 female=336	100 to 1000 mg/d 2 to 12 weeks	-BW, -WC, -WHR, -BMI
<i>Luteolin</i>	peppers, carrots, cucumber, pomegranate, herbal spices, cabbage, broccoli, medicinal herbs (sage) ³⁵			
(36) C57BL/6	DI	male n=8 per group	0.01% w/w in HFD 12 weeks	\downarrow BW, \downarrow sWAT mass, \downarrow vWAT mass, \downarrow BAT mass \downarrow eWAT adipocyte size, \uparrow IS \downarrow mast cell infiltration eWAT plasma -insulin, \downarrow leptin, \uparrow adiponectin

(37) C57BL/6	DI	male n=13 per group	0.005% w/w in HFD 16 weeks	↓BW, ↓sWAT mass, ↓vWAT mass plasma ↓TAG, ↓C, ↓NEFA, ↑IS ↑eWAT FAO genes (PGC1 α , ADRB3, CPT2, PNP2, ACAD)
(38) C57BL/6	DI	male n=8 per group	0.01% w/w in HFD 20 weeks	↓BW, ↑IS ↓plasma MCP1, IL6, TNF α ↓macrophage infiltration vWAT, ↓M1/M2-ratio
(39) C57BL/6	DI	male n=12 per group	0.01% w/w in HFD 12 weeks	↓BW, ↓weight gain, ↓fat mass ↑VO ₂ , ↑CO ₂ , ↑RER, ↑BAT UCP1 protein ↑UCP1+ cells in sWAT, ↑thermogenic genes (PGC1 α , UCP1, SIRT1, PPAR α , ELOVL3) ↑SIRT1, pAMPK, pACC protein in BAT, sWAT
(39) primary subcutaneous and brown adipocytes	acute	unknown	24 hours of 100 nM luteolin on differentiated cells	↑SIRT1, UCP1, PGC1 α protein ↑pAMPK, ↑pACC ↑thermogenic genes (UCP1, PRDM16, ELVOL6, PPAR α) ➤ effects AMPK-dependent
Catechins	grapes, apples, strawberries, apricots, broad beans, cocoa-products, green/black/oolong tea ^(40,41)			
(42) Sprague-Dawley rat	DI	male n=8 per group	2% w/w green tea extract in HFD, 2 weeks	-BW, ↓fat mass, ↑BAT weight ↑BAT DNA/protein content ↑EE, propranolol prevented ↑EE
(43) Sprague-Dawley rat	DI	male n=8 per group	0.5% w/w catechins in chow 8 weeks	-BW ↑BAT mass, ↓pWAT mass, ↓eWAT mass ↑BAT UCP1 expression plasma ↓TAG, GLC, leptin
(44) New Zealand black mice	gavage short-term	male n=6 per group	3x 500 mg/kg EGCG or placebo, chow diet	-BW, -fat mass -EE, ↓RER (p=0.053), -activity

(44) New Zealand black mice	DI	male n=11 per group	0.1% w/w EGCG in HFD DIO 4 weeks, DI 4 weeks	↓BW, ↓fat mass, -food intake, ↓eWAT weight plasma ↓TAG, -NEFA -UCP1 mRNA in BAT
(45) iBAT depots from Sprague-Dawely rat	acute	male	100 or 200 μM green tea extract for 40-90 min	↑iBAT respiration (100 μM) ↑norepinephrine (0.1 μM) stimulated respiration at 100 or 200 μM
(46) healthy men Geneva, CH	placebo crossover	male n=10	3x daily capsule with 50mg caffeine and 80 mg EGCG, 50 mg caffeine or placebo	↑diurnal EE, ↑total EE, -nocturnal EE ↓total, diurnal and nocturnal RER ↑FO, ↑urinary norepinephrine excretion
(47) young, healthy subjects Lausanne, CH	placebo crossover	male n=15 female n=16	3x daily beverage with 100 mg caffeine and 180 mg catechins or placebo, 3 days	↑total EE, diurnal EE, nocturnal EE -substrate oxidation -catecholamine secretion
(48) healthy men (BMI 23-27 kg/m ²) Laval University, CA	placebo crossover	male n=14	3x daily capsule with 200 mg caffeine plus 90, 200, 300 or 400 mg EGCG or placebo	↑total EE, -SEE -RER,-FO -catecholamine secretion
(49) overweight/obese men (BMI=31 kg/m ²) Berlin, DE	placebo crossover	male n=10	300 or 600 mg EGCG or placebo capsule for 3 days	-EE (pre- and post-meal) ↓post-meal RQ, ↑post-meal FO (300 mg), ↓post-meal CHO (300 mg) plasma -NEFA,-insulin, GLC
(50) Meta-analysis effect of EGCG on EE or anthropometric measures	8 RTC	n=268	EGCG: 300 or 600 mg/d for 2-3 days 300 to 800 mg/d for 2-12 weeks	↑EE, ↓RER, -FO, - BMI, ↓WC, -fat percentage
(51) healthy men Japanese	placebo crossover	male n=15 low BAT activity (mean SUV=1.9)	615 mg catechins plus 77 mg caffeine or placebo (81 mg caffeine), 2x daily as beverage for 5 weeks	-BMI, -fat mass, -WC, -EE ↑cold-induced thermogenesis, ↑cold-induced FO

(51) healthy men Japanese	acute crossover	male n=15	615 mg catechins plus 77 mg caffeine or placebo (81 mg caffeine), single beverage	↑post-drink EE, ↑EE in high BAT (SUV>2) vs low BAT subjects, pre-assessed by 2 hours cold-exposure
(52) healthy university students Japanese	placebo parallel	female catechin n=10 placebo n=11	640 mg catechins/d or placebo, beverage 12 weeks	-BMI, -fat mass, -BW ↑BAT density in supraclavicular region neg. correlation between EMCL and BAT density
(53) overweight/obese children Japanese	placebo parallel	catechin group (m21, f5) placebo group (m13, f6)	576 mg/d catechins or placebo (75 mg/d catechins), as Oolong tea, 12weeks	-no changes in anthropometric or metabolites in catechin vs control ↓WC, ↓SBP, ↓LDLC in catechin group when stratified to baseline values
(54) normal to overweight men Japanese ⁽³⁶⁾	placebo parallel	male catechin group n=17 placebo group n=19	690 mg/d catechins or placebo (22 mg/d), as Oolong tea, 12weeks	↓WC, ↓skinfold thickness, ↓total fat area ↓visWAT and sWAT area plasma -NEFA, -TAG, -C, -GLC, -insulin
(55) obese adult Thais (BMI>25kg/m ²)	placebo parallel	catechin group: (m21, f9) placebo group: (m21, f9)	3x daily 250 mg catechins or placebo in capsule 12 weeks	↓BMI, ↓BW, ↓fat mass, ↓WC, -HC ↓RER, ↑REE
(56) overweight/obese adults (BMI >25-32 kg/m ²) Caucasian	before-after	female n=63 male n=7	270 mg/d EGCG in capsule	-BW, ↓WC -SBP, -DBP plasma -C
(57) Meta-analysis effect of green tea extracts on anthropometry	15 RTCs	n=1243	catechin intake combined with caffeine intake (141 up to 1207 mg/d) 8 to 24 weeks	↓BMI, ↓BW, ↓WC, -WHR when compared to caffeine-intake only
Phytoestrogens	kidney beans, mung bean sprouts, Japanese arrowroot, soybean, soy products (tofu, soy milk, soy flour, soy sauce) ⁽⁵⁸⁾			

(59) C57/B6J mice	DI	male n=7-8	5% isoflavone-rich fraction of <i>Puerariae</i> flower in HFD, 7 weeks	↓BW, ↓WAT mass, ↓BAT mass -food intake, -fecal lipid content ↑VO ₂ , -RER, ↑UCP1+ cells in BAT sections
(60) CD-1 mice	DI	male n=12 female n=12	25% w/w soy-rich diet (150 ppm daidzein, 190 ppm genistein) vs soy-free diet 16 weeks	↓BW, ↓intra-abdominal fat mass, ↓iWAT, ↓eWAT/ovWAT, ↓WA adipocyte size ↓BAT mass (male), ↑brown appearance, ↓lipid droplet size ↑cold-resistance (rectal T), ↑VO ₂ , ↓RER (only male data available)
(61) Sprague-Dawley rats	ovx DI	female n=10 per group	isoflavone-rich (200 μg/g) or isoflavone-free diet 13 days	↓BW gain, ↓abdominal fat mass ↓serum leptin
(62) Long-Evans rats	DI	male	isoflavone-rich (600 ppm) or isoflavone-free (10-15 ppm) diet up to 75 days of age	↑food intake, ↓BW gain, ↓WAT mass, ↓BAT mass plasma ↑T3, ↓insulin, leptin ↑UCP1 protein in BAT
(63) Wistar rats	DIO with DI	male n=16 per group	50 mg/kg BW daidzein or vehicle, i.p. DIO 10 weeks, 14 d treatment	↓caloric intake, ↓BW gain ↓hepatic liver content plasma ↓TAG, -C, ↑GLC, -ALT ↑UCP1 protein in BAT
(64) ICR mice	DIO with gavage	male	0, 25, 50 or 100 mg/kg BW DIO 8 weeks, 30 d treatment	↓BW, ↓vWAT mass, ↓sWAT mass plasma ↓C, ↓LDL, ↓NEFA, -TAG, ↑HDL
(64) primary adipocytes differentiated from eWAT SVF	acute	male	0, 1, 3, 16, 64 μM daidzein 24 hours	↑glycerol release (dose-dependent)

(65) adipocytes from Wistar rats	acute	male	0.01, 0.1 or 1 mM daidzein	↑basal lipolysis (dose-dependent) ↑epinephrine-stimulated lipolysis (0.1 mM) ↓lipogenesis from GLC (0.1 and 1 mM)
(66) C57BL/6	DI	female n=8 per group	0.25% w/w genistein in HFD 8 weeks	↓BW, -sWAT weight, -vWAT weight ↓BAT weight (ns), ↑IS plasma -TAG, -C, HDL-, ↓LDL, ↓NEFA ↑iWAT browning (UCP1, CIDEA mRNA) ↑ hypothalamic UNC3 mRNA
(67) C57BL/6	DI	male n=7-8 per group	0.2% w/w genistein in casein diet or casein only (control) 60 days	-BW, ↑glucose tolerance ↑thermogenic genes in sWAT (UCP1, PGC1α) ↑UCP1 protein in sWAT, -BAT ↑EE, ↑VO ₂ , ↑cold-resistance (rectal T), -RER plasma -TAG, ↓GLC, ↓insulin
(67) primary adipocytes from iWAT of mice	acute	unknown	0, 5, 15, 30 μM genistein for 1 hour	-basal respiration ↑maximal respiration
(68) immortalized brown adipocytes	long-term	unknown	0, 0.1, 1 or 40 μM of genistein on differentiated adipocytes, 3 days treatment	↑UCP1 promoter activity (luciferase) ↑UCP1 activity (immunofluorescence intensity)
(69) C57BL/6	oral gavage	male and female	50 to 200 mg/kg BW genistein or vehicle for 15 d	↓BAT mass, ↓eWAT (m), ↓abdominal WAT (f) plasma ↓TAG, ↓C for 50 mg/kg BW
(70) postmenopausal women (BMI=23.6 kg/m ²) Chinese, equol-producer	placebo parallel	female n=90 per group	40 g soy flour, 40 g low-fat milk powder with 63 mg daidzein, 40 g low fat milk powder (placebo) daily, 6 months	-BW, -BMI, -WC, -HC, -WHR, -fat mass

(71) adolescent males Tasmania	placebo parallel	male isoflavone n=69 placebo n=59	50 mg isoflavone equivalents or placebo tablets daily 6 weeks	-BW
(72) obese women (20-65 yrs) (BMI 30-40 kg/m ²) USA	placebo parallel	female soy group n=22 casein group n=21	3x daily soy (50 mg isoflavone) or casein (3.5 mg isoflavone) shake, 16 weeks	-WC, -weight loss, -fat mass, -truncal fat -SBP, -DBP
(73) impaired glycemic control Chinese women (30-70 yrs)	placebo parallel	female daidzein n=55 genistein n=56 placebo n=54	10 g soy protein with no addition, 50 mg daidzein or 50 mg genistein 24 weeks	-BMI, -WC, -fat mass -IS
(74) patients with NAFLD Iranian (16-69 yrs)	placebo parallel	genistein group (m30, f11) placebo group (m31, f10)	250 mg daidzein or placebo capsules 8 weeks	-BW, ↓fat percentage, ↓WHR, ↓WC, -BMI plasma ↓TAG, -C, -LDL, -HDL, ↓insulin ↓HOMA-IR
(75) postmenopausal women (BMI>30 kg/m ²) Caucasian or AA	placebo parallel	soy group (n=17) 8 AA, 9 Caucasian placebo (n=16) 8 AA, 8 Caucasian	soy protein with isoflavones (160 mg) or placebo casein, shake 3 months	-BW, -total fat, -lean mass ↓abdominal, ↓subcutaneous abdominal fat, ↓vWAT for AA: weight loss more than for Caucasian for Caucasian: vWAT loss bigger than for AA plasma ↓IL6, -CRP, -TNFα, -leptin, -HDL, -LDL, -C, -TAG
(76) postmenopausal women (mean BMI=30.5) Caucasian	placebo parallel	female soy group n=9 placebo n=6	soy protein with isoflavones (160 mg) or placebo casein, shake 3 months	-BMI, -BW, -total fat mass, -IS ↓subcutaneous abdominal fat, ↓intra-abdominal fat plasma -GLC, -insulin

(77) Meta-analysis Effect of soy-isoflavones on BW in non-Asian, postmenopausal women	9 RCTs	isoflavones n=272 placebo n=256	40 to 160 mg/d of isoflavones 8 weeks to 1 year	↓BW with isoflavone intake <100 mg or <6 months more effective more effective with BMI<30 kg/m ²
(78) Meta-analysis effect of soy and isoflavones on anthropometric measures	24 soy RCTs 17 isoflavones RCTs	soy: f1265, m45 (1 RTC) m/f =74 (mixed) isoflavones: f1177, m0	soy protein: 7.5 to 116 mg/d 4 weeks to 2 years isoflavones: 33.3 to 300 mg/d 8 weeks to 2 years	soy: -BW, >40 g/d ↑BW, 1-3 months ↑weight gain -WC, -fat mass isoflavones: ↓BMI for postmenopausal and Caucasian women <100 mg and 2-6 months more effective -fat mass, -WC
Capsaicinoids	chili, bell peppers, jalapenos, habaneros, cayenne pepper, red pepper ^(79,80)			
(81) Std ddY mice	intra-gastric tube	unknown n=6-8	vehicle, 10 mg/kg BW capsaicin or 10, 50 mg/kg BW capsiate, 2 weeks	↓BW (ns), -food intake -BAT mass, ↓eWAT for capsiate, ↓pWAT for capsaicin and 50 mg/kg capsiate
(81) Std ddY mice	intra-gastric tube, acute	unknown n=6-8	Vehicle, 10 mg/kg BW capsaicin or 10 mg/kg BW capsiate, 3 hours	↑VO ₂ for capsaicin and capsiate, ↑serum adrenaline plasma ↑NEFA, ↓TAG
(82) C57BL/6 or TRPV1 -/- mice	intra-gastric tube, acute	male n=5-18	vehicle, 10 mg/kg BW capsaicin or 10 mg/kg BW capsiate, 3 hours	↑VO ₂ (capsaicin, capsiate at 10 mg/kg BW) ↑FO (capsaicin, capsiate at 10 mg/kg BW), ↓CHO ↑BAT and colonic T (50 mg/kg capsinoids, 10 mg/kg capsaicin) ↓T increase after denervation of jejunal nerves at 50 mg/kg capsinoid - effects in wt but not TRPV-/- mice
(83) TRPV1 -/- or wt mice	DI	male	0.01% w/w capsaicin in HFD 32 weeks	↓weight gain, ↓BW, ↑BAT UCP1, BMP8b protein

B6.129X1				<p>↑activity, ↑RER, -food intake, ↓BAT TAG content, ↑BAT glycerol release (basal or forskolin-stimulated) ↑TRPV1 protein in BAT ↑Ca²⁺ influx in isolated BA (2 μM CAP) ↑pAMPK, pSIRT1 in BAT ↓PRDM16, PPARγ acetylation (HEK293 1 μM CAP), ↑PRDM16 and PPARγ interaction in BAT lysate ➤ effects blunted in TRPV^{-/-} vs wt mice</p>
(84) TRPV1 ^{-/-} or wt mice B6.129X1	DI	male n=40 per group	0.01% w/w capsaicin in HFD 26 weeks	<p>↓weight gain, ↑TRPV1 mRNA in iWAT, eWAT ↑EE, ↑activity, -fecal lipid content, ↑RER, ↑VO₂ ↑UCP1, BMP8b, PPARα/γ protein in s/eWAT, ↑sWAT lipolysis (basal, forskolin-stimulated) ↓PRDM16, PPARγ acetylation in sWAT ↑pAMPk, CaMKKII activation in sWAT ↑Ca²⁺ influx in isolated WA (2 μM CAP) ➤ effects blunted in TRPV^{-/-} vs wt mice</p>
(85) C57BL/6	DIO+DI	male n=6 per group	0.01% w/w capsaicin in HFD DIO 10 weeks, 10 week DI	<p>↓BW, ↓weight gain, ↑eWAT, ↓rWAT mass -food intake, ↓WAT adipocyte size ↑glucose tolerance, ↑adiponectin, ↓leptin ↑TRPV1 expression WAT</p>
(86) Sprague-Dawley rats	intra-muscular	female n=9-18 per group	0.6, 0.7, 0.8 mg/kg BW capsaicin or DMSO, 80 to 120 min	<p>↑BAT and rectal T with 0.8 mg/kg BW -BAT weight, -mito content ↑BAT respiration</p>
(87) Std ddY mice	intra-gastric tube	male n=9-10	10 mg/kg BW capsiate or vehicle, 2 weeks	<p>-BAT mass, ↓eWAT, ↓pWAT ↑VO₂, ↑FO, ↑CHO, ↑UCP1 protein (BAT) ↑UCP1 mRNA (eWAT, BAT),</p>

(87) Std ddY mice	intra-gastric tube, acute	male n=4 per group	10 mg/kg BW capsiate or vehicle for 30 min	↑UCP1 mRNA in BAT
(88) healthy young men British	acute	male	breakfast with 3g chili sauce	↑post-meal EE
(89) long distance runners Japanese	acute crossover	male n=8	breakfast with or without 10 g red pepper	↑post-meal EE (30 min) ↑RER, ↑CHO, ↓FO ➤ effect blocked by propranolol
(90) healthy young men Caucasian	placebo crossover	male n=10	appetizer with or without 6 g of red pepper	↓energy intake at lunch and dinner ↑sympathetic: parasympathetic nerve activity
(91) healthy lean subjects Caucasian	placebo crossover	male n=11 female n=19	1030 mg red pepper in lunch	-EE post-meal -RER -CHO, -FO, ↑peak plasma GLP-1,-ghrelin
(92) overweight subjects (mean BMI=29.4 kg/m ²) Caucasian, AA, other	placebo parallel	male (ethnicity) Placebo (16,9,3) 3 mg (15, 9, 1) 9 mg (10, 4, 2)	0, 3, 9 mg dihydrocapsiate in gel capsule 4 weeks	-RMR (p=0.054 for 3 mg vs placebo) ↑RMR dihydrocapsiate vs placebo
(93) Meta-analysis effect of capsaicin or capsiate on EE, RER	capsiate on EE 13 RCTs capsiate on RER 9 RCTs capsaicin on EE 13 RCTs capsaicin on RER 10 RCTs female/male unknown		capsaicin doses: <7, 20-35 mg, 135-150 mg/d dihydrocapsiate doses: <1.5, 2-4 or 6-9 mg/d	↑EE at 2-9 mg/d for capsiate ↑EE at 135-150 mg/d for capsaicin ↑FO at 6-9 mg/d capsiate ↑FO at 20-150 mg/d for capsaicin ↑SNS activity (1 RCT)
(94) healthy adult participants (BMI 20-30) Caucasian	acute controlled	N=15 per group (m8, f7)	0 or 7.68 mg/d capsaicin with 100% or 75% of daily energy requirements 26 hours	-TEE 100% CAP vs 100% control (c) -DIT 75% CAP vs 100% c, ↓75 % c vs 100% -REE 75% CAP vs 100% c, ↓75 % c vs 100% ↑FAO 75% CAP vs 100% c, -75 % c vs 100% ↓CHO 75% CAP vs 100% c

(95) healthy adults BMI 25-30 kg/m ² 44% Hispanic, 41% white, non-Hispanic, 13% black, 2% others	placebo parallel	capsinoids (m21, f22) Placebo (m17, f20)	6 mg capsinoids or placebo capsule with MCTG, rapeseed oil 12 weeks	-BW, -fat mass, -abdominal fat -EE (only m): 54 kcal/d higher with CAP (p=0.19) -FO (only m): 21 mg/min higher with CAP (p=0.06)
(96) lean, healthy subjects Singapore	crossover cold vs capsinoids	Male n=8, female n=12 BAT+ m6/f6 BAT- m2/f6	12 mg capsinoids in capsule with rapeseed oil and MCTG	↑FDG-uptake in BAT ↑EE, higher in BAT+ subjects ↑FO, ↓RER plasma -GLC, -TAG, ↑C, ↑NEFA
(97) young healthy men Japanese	placebo crossover acute	male n=18 BAT+ n=10 BAT- n=8	9 mg capsinoids or placebo capsule with rapeseed oil, MCTG 2 hours	↑EE in BAT+ with capsinoids vs placebo -RER, -skin T
<i>Berberine</i>	barberry, supplements from bark, root, stems or leaves from plants of the <i>Berberis</i> genus (e.g. goldenseal, goldthread, Oregon grape, tree turmeric) ⁽⁹⁸⁾			
(99) db/db mice	i.p.	male BBR n=17 vehicle n=16	5 mg/kg BW on chow diet 26 days	↓BW, ↓eWAT mass, ↓sWAT adipocyte size ↓intra-abdominal fat, ↑IS, ↑BAT mRNA, PPAR α , PGC1 α ↓FAS ↓WAT mRNA FAS, PPAR γ , SREBP1c, aP2
(99) 3T3-L1 adipocytes	acute	male	5 μ g/mL BBR for 60 min	↑pAMPK, ↑pACC
(100) db/db mice	long-term i.p.	male n=5 per group	5 mg/kg BW on chow diet 26 days at 22°C or 30°C	↓BW, ↓fat mass, ↓plasma NEFA ↑rectal T, ↑VCO ₂ , ↑VO ₂ , ↑EE ↑cold-resistance (core T) ↑BAT activity (PET/CT) ↑BAT mito content, ↑oxphos, ↓BAT mass ↑BAT UCP1, PGC1 α , CPT1, pAMPK protein

				<p>↑iWAT thermogenic genes (UCP1, NRF1) ↑UCP1+ cells, ↑mito content, ↑UCP1 protein ➤ effects blunted at 30°C</p>
(101) C57BL/6	DI	male	5 mg/kg BW, i.p. on chow diet, 4 weeks	<p>↑hepatic FGF21 expression, ↑plasma FGF21 ↑iBAT mRNA (UCP1, DIO2, PRMD16)</p>
(102) C57BL/6	DIO	male n=7-8	1.5 mg/kg BW/d on HFD DIO 8 weeks, DI 6 weeks	<p>↓BW gain, ↓pWAT, iWAT mass ↑rectal T, ↑EE, ↑VO₂, ↑BAT activity, volume (PET/CT), ↑UCP1+ cells in BAT ↓BAT PRDM16 promoter methylation ➤ effects blunted in adiponectinCre AMPKα1/2 mice</p>
(102) BAT SVF cells	long-term	male	250 nM BBR during differentiation until d8	<p>↑basal and uncoupled respiration ↑BA adipogenesis (↑UCP1+ cells) ↑fatty acid oxidative BAT-specific genes ↑UCP1, PGC1α, PRDM16 protein</p>
(102) patients with NAFLD mean BMI=29 kg/m ² China	before-after	not defined	1.5 g BBR/d 1 month	<p>↓BW, ↓WC, ↓BMI, -total fat mass, ↓vWAT mass, ↓sWAT mass ↓HOMA-IR ↑BAT volume, BAT activity</p>
(103) subjects with NAFLD China	controlled parallel	LSI (m32/f30) LSI+P (m28/f32) LSI+BBR (m38/f24)	LSI+1.5g BBR/d, LSI+15 mg/d pioglitazone, LSI only 16 weeks	<p>↓hepatic fat content vs LSI, ↓BW, ↓BMI vs LSI and vs LSI+pioglitazone -HbA1c, ↓HOMA-IR vs LSI,</p>
(104) newly diagnosed diabetics no pharmacotherapy China	placebo parallel	placebo (m38, f28) BBR (m31, f21)	1.5 g BBR/d or placebo 12 weeks	<p>-BW, ↓BMI, -WHR plasma -FBG, ↓PBG, ↓HbA1c, -HOMA-IR, ↓C, - insulin, -TAG</p>

(105) type 2 diabetics with poor glycemic control China	before-after	n=48 sex not stated	1.5 g BBR/d plus prescribed diabetes medication 12 weeks	↓WC, ↓WHR, -BMI plasma ↓FBG, ↓PBG, -TAG, ↓C, ↓insulin ↓HOMA-IR
(106) obese adults Caucasian	before-after	male n=2 female n=5	1.5 g BBR/d 12 weeks	-BMI, -WHR, - fat percentage plasma -GLC,-TAG, ↓C, ↓ALT, ↓AST -plasma inflammatory markers

4

5 AA, African American, BAT, brown adipose tissue; BBR, berberine; BW, body weight; BMI, body mass index, C, cholesterol; CHO, carbohydrate
6 oxidation; d, day; DI, dietary intervention; DIO, diet-induced obesity; DIT, diet-induced thermogenesis; EE, energy expenditure; EMCL,
7 extramyocellular lipid; f, female; FBG, fasting blood glucose; FO, fat oxidation; HC, hip circumference; HFHS, high fat high sucrose; IS, insulin
8 sensitivity, IGT, impaired glucose tolerance; i.p., intraperitoneal; LSI, life style intervention; m, male; MCTG, medium chain triglyceride; mito,
9 mitochondria(l); NAFLD, non-alcoholic fatty liver disease, NEFA, non-esterified fatty acids; ovx, ovariectomized, PBG, postprandial blood glucose;
10 RCT, randomized controlled trial; REE, resting energy expenditure; RER, resting energy expenditure; SEE, sleeping energy expenditure; SUV,
11 standardized uptake value; T, temperature; TAG, triacylglycerol, TEE, total energy expenditure; WC, waist circumference; WHR, waist-hip-ratio; WAT,
12 white adipose tissue, wt, wild-type; - unchanged, ↓decrease, ↑increase compared to placebo or baseline

13 References

- 14 1. Reinisalo M, Kårlund A, Koskela A, *et al.* (2015) Polyphenol Stilbenes: Molecular
15 Mechanisms of Defence against Oxidative Stress and Aging-Related Diseases. *Oxid Med*
16 *Cell Longev* **2015**, 340520.
- 17 2. Rimando AM, Nagmani R, Feller DR, *et al.* (2005) Pterostilbene, a new agonist for the
18 peroxisome proliferator-activated receptor alpha-isoform, lowers plasma lipoproteins and
19 cholesterol in hypercholesterolemic hamsters. *J Agric Food Chem* **53**, 3403–3407.
20
- 21 3. Etxeberria U, Hijona E, Aguirre L, *et al.* (2017) Pterostilbene-induced changes in gut
22 microbiota composition in relation to obesity. *Mol Nutr Food Res* **61**, 1500906
- 23 4. Gómez-Zorita S, Fernández-Quintela A, Lasa A, *et al.* (2014) Pterostilbene, a dimethyl
24 ether derivative of resveratrol, reduces fat accumulation in rats fed an obesogenic diet. *J*
25 *Agric Food Chem* **62**, 8371–8378.
- 26 5. Aguirre L, Milton-Laskibar I, Hijona E, *et al.* (2016) Effects of pterostilbene in brown
27 adipose tissue from obese rats. *J Physiol Biochem* **73**, 457–464.
- 28 6. Nagao K, Jinnouchi T, Kai S, *et al.* (2017) Pterostilbene, a dimethylated analog of
29 resveratrol, promotes energy metabolism in obese rats. *J Nutr Biochem* **43**, 151–155.
- 30 7. La Spina M, Galletta E, Azzolini M, *et al.* (2019) Browning Effects of a Chronic
31 Pterostilbene Supplementation in Mice Fed a High-Fat Diet. *Int J Mol Sci* **20**, 5377.
- 32 8. Riche DM, Riche KD, Blackshear CT, *et al.* (2014) Pterostilbene on metabolic
33 parameters: a randomized, double-blind, and placebo-controlled trial. *Evid.-Based*
34 *Complement Alter Med ECAM* **2014**, 459165.
35
- 36 9. Burns J, Yokota T, Ashihara H, *et al.* (2002) Plant foods and herbal sources of
37 resveratrol. *J Agric Food Chem* **50**, 3337–3340.
- 38 10. Lagouge M, Argmann C, Gerhart-Hines Z, *et al.* (2006) Resveratrol improves
39 mitochondrial function and protects against metabolic disease by activating SIRT1 and
40 PGC-1alpha. *Cell* **127**, 1109–1122.
- 41 11. Baur JA, Pearson KJ, Price NL, *et al.* (2006) Resveratrol improves health and survival
42 of mice on a high-calorie diet. *Nature* **444**, 337–342.
- 43 12. Dal-Pan A, Blanc S & Aujard F (2010) Resveratrol suppresses body mass gain in a
44 seasonal non-human primate model of obesity. *BMC Physiol* **10**, 11.
- 45 13. Andrade JMO, Frade ACM, Guimarães JB, *et al.* (2014) Resveratrol increases brown
46 adipose tissue thermogenesis markers by increasing SIRT1 and energy expenditure and
47 decreasing fat accumulation in adipose tissue of mice fed a standard diet. *Eur J Nutr* **53**,
48 1503–1510.
- 49 14. Wang S, Liang X, Yang Q, *et al.* (2015) Resveratrol induces brown-like adipocyte
50 formation in white fat through activation of AMP-activated protein kinase (AMPK) α 1.
51 *Int J Obes* **2005** **39**, 967–976.

- 52 15. Wang S, Liang X, Yang Q, *et al.* (2017) Resveratrol enhances brown adipocyte formation
53 and function by activating AMP-activated protein kinase (AMPK) $\alpha 1$ in mice fed high-
54 fat diet. *Mol Nut Food Res* **61**, 1600746.
- 55 16. Alberdi G, Rodríguez VM, Miranda J, *et al.* (2013) Thermogenesis is involved in the
56 body-fat lowering effects of resveratrol in rats. *Food Chem* **141**, 1530–1535.
- 57 17. Poulsen MM, Vestergaard PF, Clasen BF, *et al.* (2013) High-Dose Resveratrol
58 Supplementation in Obese Men. *Diabetes* **62**, 1186–1195.
- 59 18. Yoshino J, Conte C, Fontana L, *et al.* (2012) Resveratrol supplementation does not
60 improve metabolic function in nonobese women with normal glucose tolerance. *Cell*
61 *Metab* **16**, 658–664.
- 62 19. Crandall JP, Oram V, Trandafirescu G, *et al.* (2012) Pilot study of resveratrol in older
63 adults with impaired glucose tolerance. *J Gerontol A Biol Sci Med Sci* **67**, 1307–1312.
- 64 20. Timmers S, Konings E, Bilet L, *et al.* (2011) Calorie restriction-like effects of 30 days
65 of Resveratrol (resVida™) supplementation on energy metabolism and metabolic profile
66 in obese humans. *Cell Metab* **14**, 612–622.
- 67 21. Li Y, Yao J, Han C, *et al.* (2016) Quercetin, Inflammation and Immunity. *Nutrients* **8**,
68 187
69
- 70 22. Moon J, Do H-J, Kim OY, *et al.* (2013) Antiobesity effects of quercetin-rich onion peel
71 extract on the differentiation of 3T3-L1 preadipocytes and the adipogenesis in high fat-
72 fed rats. *Food Chem Toxicol Int J Publ Br Ind Biol Res Asso* **58**, 347–354.
- 73 23. Ting Y, Chang W-T, Shiau D-K, *et al.* (2018) Antiobesity Efficacy of Quercetin-Rich
74 Supplement on Diet-Induced Obese Rats: Effects on Body Composition, Serum Lipid
75 Profile, and Gene Expression. *J Agric Food Chem* **66**, 70–80.
- 76 24. Rivera L, Morón R, Sánchez M, *et al.* (2008) Quercetin ameliorates metabolic syndrome
77 and improves the inflammatory status in obese Zucker rats. *Obes. Silver Spring Md* **16**,
78 2081–2087.
- 79 25. Lee SG, Parks JS & Kang HW (2017) Quercetin, a functional compound of onion peel,
80 remodels white adipocytes to brown-like adipocytes. *J Nutr Biochem* **42**, 62–71.
- 81 26. Kuipers EN, van Dam AD, Held NM, *et al.* (2018) Quercetin Lowers Plasma
82 Triglycerides Accompanied by White Adipose Tissue Browning in Diet-Induced Obese
83 Mice. *Int J Mol Sci* **19**.
- 84 27. Stewart LK, Soileau JL, Ribnicky D, *et al.* (2008) Quercetin transiently increases energy
85 expenditure but persistently decreases circulating markers of inflammation in C57BL/6J
86 mice fed a high-fat diet. *Metabolism* **57**, S39–S46.
- 87 28. Choi H, Kim C-S & Yu R (2018) Quercetin Upregulates Uncoupling Protein 1 in
88 White/Brown Adipose Tissues through Sympathetic Stimulation. *J Obes Metab Syndr*
89 **27**, 102–109.

- 90 29. Kuppusamy UR & Das NP (1994) Potentiation of beta-adrenoceptor agonist-mediated
91 lipolysis by quercetin and fisetin in isolated rat adipocytes. *Biochem Pharmacol* **47**, 521–
92 529.
- 93 30. Dong J, Zhang X, Zhang L, *et al.* (2014) Quercetin reduces obesity-associated ATM
94 infiltration and inflammation in mice: a mechanism including AMPK α 1/SIRT1. *J Lipid*
95 *Res* **55**, 363–374.
- 96 31. Kim J, Cha Y-J, Lee K-H, *et al.* (2013) Effect of onion peel extract supplementation on
97 the lipid profile and antioxidative status of healthy young women: a randomized,
98 placebo-controlled, double-blind, crossover trial. *Nutr Res Pract* **7**, 373–379.
- 99 32. Kim K-A & Yim J-E (2016) The Effect of Onion Peel Extract on Inflammatory
100 Mediators in Korean Overweight and Obese Women. *Clin Nutr Res* **5**, 261–269.
- 101 33. Lee J-S, Cha Y-J, Lee K-H, *et al.* (2016) Onion peel extract reduces the percentage of
102 body fat in overweight and obese subjects: a 12-week, randomized, double-blind,
103 placebo-controlled study. *Nut Res Pract* **10**, 175–181.
- 104 34. Huang H, Liao D, Dong Y, *et al.* (2019) Clinical effectiveness of quercetin
105 supplementation in the management of weight loss: a pooled analysis of randomized
106 controlled trials. *Diabetes Metab Syndr Obes Targets Ther* **12**, 553–563
107
- 108 35. López-Lázaro M (2009) Distribution and biological activities of the flavonoid luteolin.
109 *Mini Rev Med Chem* **9**, 31-59
110
- 111 36. Xu N, Zhang L, Dong J, *et al.* (2014) Low-dose diet supplement of a natural flavonoid,
112 luteolin, ameliorates diet-induced obesity and insulin resistance in mice. *Mol Nutr Food*
113 *Res* **58**, 1258–1268.
- 114 37. Kwon E-Y, Jung UJ, Park T, *et al.* (2015) Luteolin attenuates hepatic steatosis and insulin
115 resistance through the interplay between the liver and adipose tissue in mice with diet-
116 induced obesity. *Diabetes* **64**, 1658–1669.
- 117 38. Zhang L, Han Y-J, Zhang X, *et al.* (2016) Luteolin reduces obesity-associated insulin
118 resistance in mice by activating AMPK α 1 signalling in adipose tissue macrophages.
119 *Diabetologia* **59**, 2219–2228.
- 120 39. Zhang X, Zhang Q-X, Wang X, *et al.* (2016) Dietary luteolin activates browning and
121 thermogenesis in mice through an AMPK/PGC1 α pathway-mediated mechanism. *Int J*
122 *Obes* **40**, 1841–1849.
- 123 40. Henning SM, Fajardo-Lira C, Lee HW, *et al.* (2003) Catechin Content of 18 Teas and a
124 Green Tea Extract Supplement Correlates With the Antioxidant Capacity. *Nutr Cancer*
125 **45**, 226–235.
- 126 41. Arts ICW, van de Putte B & Hollman PCH (2000) Catechin Contents of Foods
127 Commonly Consumed in The Netherlands. 2. Tea, Wine, Fruit Juices, and Chocolate
128 Milk. *J. Agric Food Chem.* **48**, 1752–1757.

- 129 42. Choo JJ (2003) Green tea reduces body fat accretion caused by high-fat diet in rats
130 through beta-adrenoceptor activation of thermogenesis in brown adipose tissue. *J. Nutr*
131 *Biochem* **14**, 671–676.
- 132 43. Nomura S, Ichinose T, Jinde M, *et al.* (2008) Tea catechins enhance the mRNA
133 expression of uncoupling protein 1 in rat brown adipose tissue. *J Nutr. Biochem* **19**, 840–
134 847.
- 135 44. Klaus S, Pültz S, Thöne-Reineke C, *et al.* (2005) Epigallocatechin gallate attenuates diet-
136 induced obesity in mice by decreasing energy absorption and increasing fat oxidation.
137 *Int J Obes* 2005 **29**, 615–623.
- 138 45. Dulloo AG, Seydoux J, Girardier L, *et al.* (2000) Green tea and thermogenesis:
139 interactions between catechin-polyphenols, caffeine and sympathetic activity. *Int J Obes*
140 *Relat Metab Disord J Int Assoc Study Obes* **24**, 252–258.
- 141 46. Dulloo AG, Duret C, Rohrer D, *et al.* (1999) Efficacy of a green tea extract rich in
142 catechin polyphenols and caffeine in increasing 24-h energy expenditure and fat
143 oxidation in humans. *Am. J. Clin. Nutr* **70**, 1040–1045.
- 144 47. Rudelle S, Ferruzzi MG, Cristiani I, *et al.* (2007) Effect of a thermogenic beverage on
145 24-hour energy metabolism in humans. *Obes Silver Spring Md* **15**, 349–355.
- 146 48. Bérubé-Parent S, Pelletier C, Doré J, *et al.* (2005) Effects of encapsulated green tea and
147 Guarana extracts containing a mixture of epigallocatechin-3-gallate and caffeine on 24 h
148 energy expenditure and fat oxidation in men. *Br J Nutr* **94**, 432–436.
- 149 49. Thielecke F, Rahn G, Böhnke J, *et al.* (2010) Epigallocatechin-3-gallate and postprandial
150 fat oxidation in overweight/obese male volunteers: a pilot study. *Eur J Clin Nutr* **64**,
151 704–713.
- 152 50. Kapoor MP, Sugita M, Fukuzawa Y, *et al.* (2017) Physiological effects of
153 epigallocatechin-3-gallate (EGCG) on energy expenditure for prospective fat oxidation
154 in humans: A systematic review and meta-analysis. *Nutr Biochem* **43**, 1–10.
- 155 51. Yoneshiro T, Matsushita M, Hibi M, *et al.* (2017) Tea catechin and caffeine activate
156 brown adipose tissue and increase cold-induced thermogenic capacity in humans. *Am. J*
157 *Clin Nutr* **105**, 873–881.
- 158 52. Nirengi S, Amagasa S, Homma T, *et al.* (2016) Daily ingestion of catechin-rich beverage
159 increases brown adipose tissue density and decreases extramyocellular lipids in healthy
160 young women. *SpringerPlus* **5**, 1363.
- 161 53. Matsuyama T, Tanaka Y, Kamimaki I, *et al.* (2008) Catechin safely improved higher
162 levels of fatness, blood pressure, and cholesterol in children. *Obes Silver Spring Md* **16**,
163 1338–1348.
- 164 54. Nagao T, Komine Y, Soga S, *et al.* (2005) Ingestion of a tea rich in catechins leads to a
165 reduction in body fat and malondialdehyde-modified LDL in men. *Am J Clin Nut.* **81**,
166 122–129.

- 167 55. Auvichayapat P, PrapoChanung M, Tunkamnerdthai O, *et al.* (2008) Effectiveness of
168 green tea on weight reduction in obese Thais: A randomized, controlled trial. *Physiol*
169 *Behav* **93**, 486–491.
- 170 56. Chantre P & Lairon D (2002) Recent findings of green tea extract AR25 (Exolise) and
171 its activity for the treatment of obesity. *Phytomedicine* **9**, 3–8.
- 172 57. Phung OJ, Baker WL, Matthews LJ, *et al.* (2010) Effect of green tea catechins with or
173 without caffeine on anthropometric measures: a systematic review and meta-analysis.
174 *Am J Clin Nutr* **91**, 73–81.
- 175 58. Zaheer K & Humayoun Akhtar M (2017) An updated review of dietary isoflavones:
176 Nutrition, processing, bioavailability and impacts on human health. *Crit Rev Food Sci*
177 *Nutr* **57**, 1280–1293.
- 178 59. Kamiya T, Nagamine R, Sameshima-Kamiya M, *et al.* (2012) The isoflavone-rich
179 fraction of the crude extract of the Puerariae flower increases oxygen consumption and
180 BAT UCP1 expression in high-fat diet-fed mice. *Glob J Health Sc.* **4**, 147–155.
- 181 60. Cederroth CR, Vinciguerra M, Kühne F, *et al.* (2007) A Phytoestrogen-Rich Diet
182 Increases Energy Expenditure and Decreases Adiposity in Mice. *Environ Health*
183 *Perspect* **115**, 1467–1473.
- 184 61. Russell AL, Grimes JM, Cruthirds DF, *et al.* (2017) Dietary Isoflavone-Dependent and
185 Estradiol Replacement Effects on Body Weight in the Ovariectomized (OVX) Rat. *Horm*
186 *Metab Res Horm Stoffwechselforschung Horm Metab* **49**, 457–465.
- 187 62. Lephart ED, Porter JP, Lund TD, *et al.* (2004) Dietary isoflavones alter regulatory
188 behaviors, metabolic hormones and neuroendocrine function in Long-Evans male rats.
189 *Nutr. Metab.* **1**, 16.
- 190 63. Crespillo A, Alonso M, Vida M, *et al.* (2011) Reduction of body weight, liver steatosis
191 and expression of stearyl-CoA desaturase 1 by the isoflavone daidzein in diet-induced
192 obesity. *Br J Pharmacol* **164**, 1899–1915
193
- 194 64. Guo Y, Wu G, Su X, *et al.* (2009) Antiobesity action of a daidzein derivative on male
195 obese mice induced by a high-fat diet. *Nutr Res N. Y. N* **29**, 656–663.
- 196 65. Szkudelska K, Szkudelski T & Nogowski L (2002) Daidzein, coumestrol and
197 zearalenone affect lipogenesis and lipolysis in rat adipocytes. *Phytomedicine Int J*
198 *Phytother Phytopharm* **9**, 338–345.
- 199 66. Zhou L, Xiao X, Zhang Q, *et al.* (2019) A Possible Mechanism: Genistein Improves
200 Metabolism and Induces White Fat Browning Through Modulating Hypothalamic
201 Expression of Ucn3, Depp, and Stc1. *Front Endocrino.* **10**, 478.
- 202 67. Palacios-González B, Vargas-Castillo A, Velázquez-Villegas LA, *et al.* (2019) Genistein
203 increases the thermogenic program of subcutaneous WAT and increases energy
204 expenditure in mice. *J Nutr Biochem* **68**, 59–68.
- 205 68. Buhlmann E, Horváth C, Houriet J, *et al.* (2019) Puerariae lobatae root extracts and the
206 regulation of brown fat activity. *Phytomedicine Int J Phytother Phytopharm* **64**, 153075.

- 207 69. Penza M, Montani C, Romani A, *et al.* (2006) Genistein affects adipose tissue deposition
208 in a dose-dependent and gender-specific manner. *Endocrinology* **147**, 5740–5751.
- 209 70. Liu Z-M, Ho SC, Chen Y-M, *et al.* (2013) A six-month randomized controlled trial of
210 whole soy and isoflavones daidzein on body composition in equol-producing
211 postmenopausal women with prehypertension. *J Obes* **2013**, 359763.
- 212 71. Jones G, Dwyer T, Hynes K, *et al.* (2003) A randomized controlled trial of phytoestrogen
213 supplementation, growth and bone turnover in adolescent males. *Eur J Clin Nut.* **57**, 324–
214 327.
- 215 72. Anderson JW, Fuller J, Patterson K, *et al.* (2007) Soy compared to casein meal
216 replacement shakes with energy-restricted diets for obese women: randomized controlled
217 trial. *Metabolism* **56**, 280–288.
- 218 73. Ye Y-B, Chen A-L, Lu W, *et al.* (2015) Daidzein and genistein fail to improve glycemic
219 control and insulin sensitivity in Chinese women with impaired glucose regulation: A
220 double-blind, randomized, placebo-controlled trial. *Mol Nut Foodes* **59**, 240–249.
- 221 74. Amanat S, Eftekhari MH, Fararouei M, *et al.* (2018) Genistein supplementation improves
222 insulin resistance and inflammatory state in non-alcoholic fatty liver patients: A
223 randomized, controlled trial. *Clin Nutr Edinb Scotl* **37**, 1210–1215.
- 224 75. Christie DR, Grant J, Darnell BE, *et al.* (2010) Metabolic effects of soy supplementation
225 in postmenopausal Caucasian and African American women: a randomized, placebo-
226 controlled trial. *Am J Obstet Gynecol* **203**, 153.e1–9.
- 227 76. Sites CK, Cooper BC, Toth MJ, *et al.* (2007) Effect of a daily supplement of soy protein
228 on body composition and insulin secretion in postmenopausal women. *Fertil Steril* **88**,
229 1609–1617.
- 230 77. Zhang Y-B, Chen W-H, Guo J-J, *et al.* (2013) Soy isoflavone supplementation could
231 reduce body weight and improve glucose metabolism in non-Asian postmenopausal
232 women--a meta-analysis. *Nutr Burbank Los Angel Cty Calif* **29**, 8–14.
- 233 78. Akhlaghi M, Zare M & Nouripour F (2017) Effect of Soy and Soy Isoflavones on
234 Obesity-Related Anthropometric Measures: A Systematic Review and Meta-analysis of
235 Randomized Controlled Clinical Trials. *Adv Nutr* **8**, 705–717.
- 236 79. Orellana-Escobedo L, Garcia-Amezquita LE, Olivas GI, *et al.* (2013) Capsaicinoids
237 content and proximate composition of Mexican chili peppers (*Capsicum* spp.) cultivated
238 in the State of Chihuahua. *CyTA - J. Food*, 179–184.
- 239 80. Scientific Committee on Food (2002) *Opinion of the Scientific Committee on Food on*
240 *Capsaicin*. European Commision Health&Consumer protection directorate-general
241 Brussel, Belgium.
- 242 81. Ohnuki K, Haramizu S, Oki K, *et al.* (2001) Administration of capsiate, a non-pungent
243 capsaicin analog, promotes energy metabolism and suppresses body fat accumulation in
244 mice. *Biosci Biotechnol Biochem* **65**, 2735–2740.

- 245 82. Kawabata F, Inoue N, Masamoto Y, *et al.* (2009) Non-pungent capsaicin analogs
246 (capsinoids) increase metabolic rate and enhance thermogenesis via gastrointestinal
247 TRPV1 in mice. *Biosci Biotechnol Biochem* **73**, 2690–2697.
- 248 83. Baskaran P, Krishnan V, Fettel K, *et al.* (2017) TRPV1 activation counters diet-induced
249 obesity through sirtuin-1 activation and PRDM-16 deacetylation in brown adipose tissue.
250 *Int. J. Obes. 2005* **41**, 739–749.
- 251 84. Baskaran P, Krishnan V, Ren J, *et al.* (2016) Capsaicin induces browning of white
252 adipose tissue and counters obesity by activating TRPV1 channel-dependent
253 mechanisms. *Br J Pharmacol* **173**, 2369–2389.
- 254 85. Kang J-H, Goto T, Han I-S, *et al.* (2010) Dietary capsaicin reduces obesity-induced
255 insulin resistance and hepatic steatosis in obese mice fed a high-fat diet. *Obes Silver*
256 *Spring Md* **18**, 780–787.
- 257 86. Yoshida T, Yoshioka K, Wakabayashi Y, *et al.* (1988) Effects of capsaicin and
258 isothiocyanate on thermogenesis of interscapular brown adipose tissue in rats. *J Nutr Sci*
259 *Vitaminol (Tokyo)* **34**, 587–594.
- 260 87. Masuda Y, Haramizu S, Oki K, *et al.* (2003) Upregulation of uncoupling proteins by oral
261 administration of capsiate, a nonpungent capsaicin analog. *J Appl Physiol Bethesda Md*
262 *1985* **95**, 2408–2415.
- 263 88. Henry CJ & Emery B (1986) Effect of spiced food on metabolic rate. *Hum Nutr Clin*
264 *Nutr* **40**, 165–168.
- 265 89. Yoshioka M, Lim K, Kikuzato S, *et al.* (1995) Effects of red-pepper diet on the energy
266 metabolism in men. *J Nutr Sci Vitaminol (Tokyo)* **41**, 647–656.
- 267 90. Yoshioka M, St-Pierre S, Drapeau V, *et al.* (1999) Effects of red pepper on appetite and
268 energy intake. *Br J Nutr* **82**, 115–123.
- 269 91. Smeets AJ & Westerterp-Plantenga MS (2009) The acute effects of a lunch containing
270 capsaicin on energy and substrate utilisation, hormones, and satiety. *Eur J Nutr* **48**, 229–
271 234.
- 272 92. Galgani JE & Ravussin E (2010) Effect of dihydrocapsiate on resting metabolic rate in
273 humans. *Am J Clin Nutr* **92**, 1089–1093.
- 274 93. Ludy M-J, Moore GE & Mattes RD (2012) The Effects of Capsaicin and Capsiate on
275 Energy Balance: Critical Review and Meta-analyses of Studies in Humans. *Chem Senses*
276 **37**, 103–121.
- 277 94. Janssens PLHR, Hursel R, Martens EAP, *et al.* (2013) Acute Effects of Capsaicin on
278 Energy Expenditure and Fat Oxidation in Negative Energy Balance. *PLoS ONE* **8**.
279 e67786
- 280 95. Snitker S, Fujishima Y, Shen H, *et al.* (2009) Effects of novel capsinoid treatment on
281 fatness and energy metabolism in humans: possible pharmacogenetic implications. *Am.*
282 *J Clin Nutr* **89**, 45–50.

- 283 96. Sun L, Camps SG, Goh HJ, *et al.* (2018) Capsinoids activate brown adipose tissue (BAT)
284 with increased energy expenditure associated with subthreshold 18-fluorine
285 fluorodeoxyglucose uptake in BAT-positive humans confirmed by positron emission
286 tomography scan. *Am J Clin Nutr* **107**, 62–70.
- 287 97. Yoneshiro T, Aita S, Kawai Y, *et al.* (2012) Nonpungent capsaicin analogs (capsinoids)
288 increase energy expenditure through the activation of brown adipose tissue in humans.
289 *Am J Clin Nutr* **95**, 845–850.
- 290 98. Neag MA, Mocan A, Echeverría J, *et al.* (2018) Berberine: Botanical Occurrence,
291 Traditional Uses, Extraction Methods, and Relevance in Cardiovascular, Metabolic,
292 Hepatic, and Renal Disorders. *Front Pharmacol* **9**, 557.
- 293 99. Lee YS, Kim WS, Kim KH, *et al.* (2006) Berberine, a natural plant product, activates
294 AMP-activated protein kinase with beneficial metabolic effects in diabetic and insulin-
295 resistant states. *Diabetes* **55**, 2256–2264.
- 296 100. Zhang Z, Zhang H, Li B, *et al.* (2014) Berberine activates thermogenesis in white and
297 brown adipose tissue. *Nat Commun* **5**, 5493.
- 298 101. Li Y, Wong K, Giles A, *et al.* (2014) Hepatic SIRT1 attenuates hepatic steatosis and
299 controls energy balance in mice by inducing fibroblast growth factor 21.
300 *Gastroenterology* **146**, 539-549.e7.
- 301 102. Wu L, Xia M, Duan Y, *et al.* (2019) Berberine promotes the recruitment and activation
302 of brown adipose tissue in mice and humans. *Cell Death Dis* **10**, 468.
- 303 103. Yan H-M, Xia M-F, Wang Y, *et al.* (2015) Efficacy of Berberine in Patients with Non-
304 Alcoholic Fatty Liver Disease. *PloS One* **10**, e0134172.
- 305 104. Zhang Y, Li X, Zou D, *et al.* (2008) Treatment of type 2 diabetes and dyslipidemia with
306 the natural plant alkaloid berberine. *J Clin Endocrinol Metab* **93**, 2559–2565.
- 307 105. Yin J, Xing H & Ye J (2008) Efficacy of berberine in patients with type 2 diabetes
308 mellitus. *Metabolism* **57**, 712–717.
- 309 106. Hu Y, Ehli EA, Kittelsrud J, *et al.* (2012) Lipid-lowering effect of berberine in human
310 subjects and rats. *Phytomedicine Int J Phytother Phytopharm* **19**, 861–867.