

**28S typical amplification**

Generate 1.1-1.3 using genomic DNA, then run 2.1-2.2 (using product from step 1.1), 2.3-2.4 (product from step 1.2) (see notes about 2.3a), and 2.5-2.6 (product from step 1.3). 2.5 may be used as positive control to detect the presence of DNA (see the respective note)

Name	1.1. 28SD1-5.P		1.2. 28SD5-10.P		1.3. 28SD11.P		2.1. 28SD1-5P.s1 (=28S1)		2.2. 28SD1-5P.n1 (=28S3L)		2.3. 28SD5-10P.n1 (=28SIV)		2.3a. 28SD5-10P.n1 (=28S4)		2.4. 28SD5-10P.n2 (=28S5)		2.5. 28SD11P.n1 (=28SD9)		2.6. 28SD11Psh2 (=28S11a)	
approx size	1100		860		810		1035		835		650		800							
Primer (F)	28S_f1_4F		28S_NS2F_m		28S_1656F		28S_1F_Arach3_T		28Sa_DD_T		28S_NS2_11Fm_T		28S_3677F		28S_NS2_21Rm_T		28S_V(D9-10)		28S_3528F_Arach3_T	
Primer (R)	28S_f1_3R		28S_NS2R		28S_3531R_Arach3		28S_1661R_T		28SIR_scar2_T		28S_NS2_11Rm_T		28S_NS2_11R_v3		28S_NS2_21R_T		28S_V(D9-10)		28S_3528R_Arach3_T	
dH <sub>2</sub> O	9.81	9.81	9.81	9.81	9.81	9.81	13.22	13.22	13.22	13.22	13.22	13.22	13.22	13.22	13.22	12.93	12.93	12.93	12.93	13.22
PCR buffer	1.5	1.5	1.5	1.5	1.5	1.5	2	2	2	2	2	2	2	2	2	2	2	2	2	2
MgSO <sub>4</sub> (50 mM)	1.05	1.05	1.05	1.05	1.05	1.05	1.4	1.4	1.4	1.4	1.4	1.4	1.4	1.4	1.4	1.4	1.4	1.4	1.4	1.4
dNTPs (10 mM each)	1.05	1.05	1.05	1.05	1.05	1.05	1.4	1.4	1.4	1.4	1.4	1.4	1.4	1.4	1.4	1.4	1.4	1.4	1.4	1.4
Primers (10uM each)	0.6	0.6	0.6	0.6	0.6	0.6	0.8	0.8	0.8	0.8	0.8	0.8	0.8	0.8	0.8	0.8	0.8	0.8	0.8	0.8
Plat Taq (1u-0.2ul- $\mu$ )	0.09	0.09	0.09	0.09	0.09	0.09	0.08	0.08	0.08	0.08	0.08	0.08	0.08	0.08	0.08	0.08	0.08	0.08	0.08	0.08
DNA	0.3	0.3	0.3	0.3	0.3	0.3	0.3	0.3	0.3	0.3	0.3	0.3	0.3	0.3	0.3	0.3	0.3	0.3	0.3	0.3
n Volumes (1 sample)	15	15	15	15	15	15	20	20	20	20	20	20	20	20	20	20	20	20	20	20
PCR Profile	°C		°C		°C		°C		°C		°C		°C		°C		°C		°C	
denature	94	0:30	94	0:30	94	0:30	94	0:30	94	0:30	94	0:30	94	0:30	94	0:30	94	0:30	94	0:30
anneal	50	0:30	51	0:35	51	0:35	51	0:35	51	0:35	51	0:35	51	0:35	51	0:35	51	0:35	51	0:35
extension	72	2:15+1s/cycle	72	2:20+1s/cycle	72	2:20+1s/cycle	72	2:20+1s/cycle	72	2:20+1s/cycle	72	2:20+1s/cycle	72	2:20+1s/cycle	72	2:20+1s/cycle	72	2:20+1s/cycle	72	2:20+1s/cycle
#cycles	35		35		35		35		35		35		35		35		35		35	
denature																				
anneal																				
extension																				
#cycles																				

**NOTE**

Primer Name	Sequence (5' to 3')	M13 tails
28S_f1_4F	GACCTCAGATCARGGcGAH	-
28S_f1_3R	GCTGTTCACATGRAACCTTC	-
28S_NS2F_m	TCTTGAACACCGGACCAAGG	-
28S_NS2R	AACCTGTCTCACGACGGTCT	-
28S_1656F	TACCVATATCCGAKCAGGT	-
28S_3531R_Arach3	AAGCYTCARTAGATCCGAT	-
28S_1F_Arach3_T	TGTAAAACGACGGCCAGTACCCGCTGAATTTAAGCAT	FORW
28S_1661R_T	CAGGAAACAGCTATGACCACTTCCGGTA	REV
28Sa_DD_T	TGTAAAACGACGGCCAGTGACCCGCTGTGAAACACGGA	FORW
28SIR_scar2_T	CAGGAAACAGCTATGACCTGCTACTACCACCAGATCTGC	REV
28S_NS2F_11Fm_T	TGTAAAACGACGGCCAGTGGCCATTTTGGTAAAGCAGA	FORW
28S_NS2_11Rm_T	CAGGAAACAGCTATGACCBTTTCCGACTTCCTTA	REV
28S_NS2_21Rm_T	TGTAAAACGACGGCCAGTATATCCGACAGGCTCTCC	FORW
28S_NS2_21R_T	CAGGAAACAGCTATGACCGTCTTCTTCCCGCTGATT	REV
28SV(D9-10)	GTAGCCAAATGCCTCTGCA	-
28SX(D9-10)	CACAATGATGGAAGGCC	-
28S_2732F_T	TGTAAAACGACGGCCAGTTGACTGGGCGGTACATC	FORW
28S_3528R_Arach3	T CAGGAAACAGCTATGACCGCTCARTAGATGCGCATGG	REV
28S_3677F	TCAAACCTTTRAAATGGGTGAGATG	-
28S_NS2_11R_v3	CCATCTTCAGAGCCAATCC	-

Use 2.3a if does not work or Alternative to 2.3 (more specific to astigmatid mites). Use if 2.3 does not work or produces seperimposed sequences  
 Because these primers are universally conserved, this protocol is used for amplification from genomic DNA as positive control to detect if DNA is present.  
 Use 34 cycles and dna=0.3 for nested amplification