Supplementary Materials

(Sall et al.)

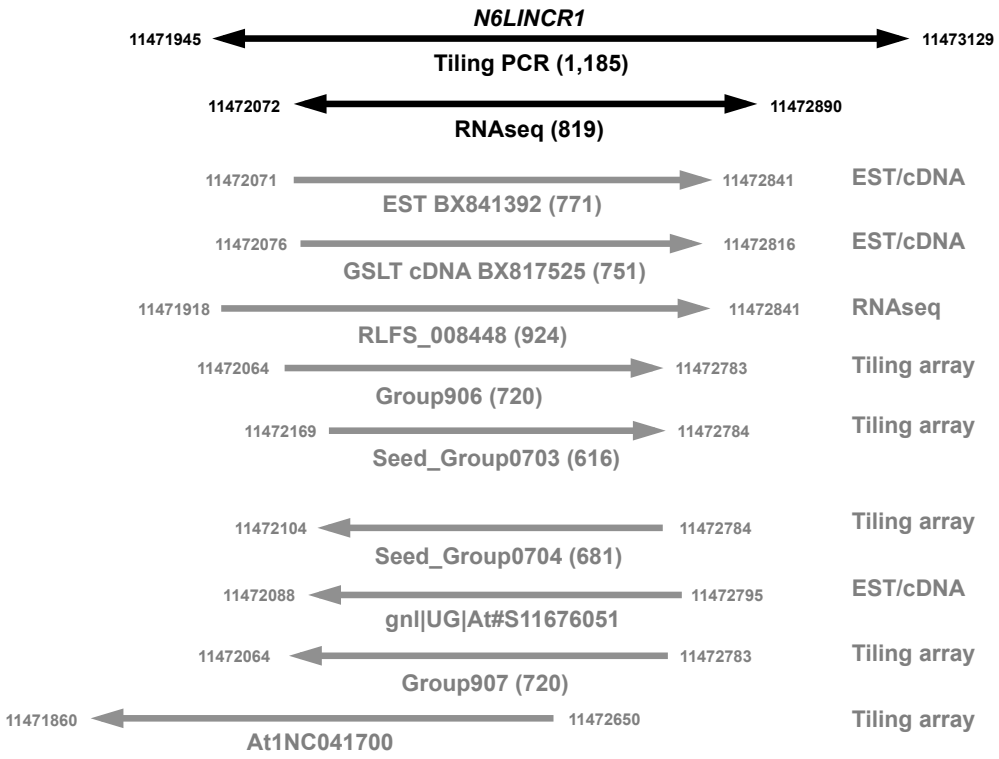


Figure S1. Transcripts detected in the previous and present studies that were originated from the *N6LINCR1* (lincRNA upregulated upon *NCED6* induction) genomic region. The length and positions of other transcripts relative to *N6LINCR1* are shown according to previous reports (Matsui et al., 2008; Nakashima et al., 2009; Okamoto et al., 2010; Richter et al., 2010; Visscher et al., 2010; Qin et al., 2011; Jin et al., 2013).

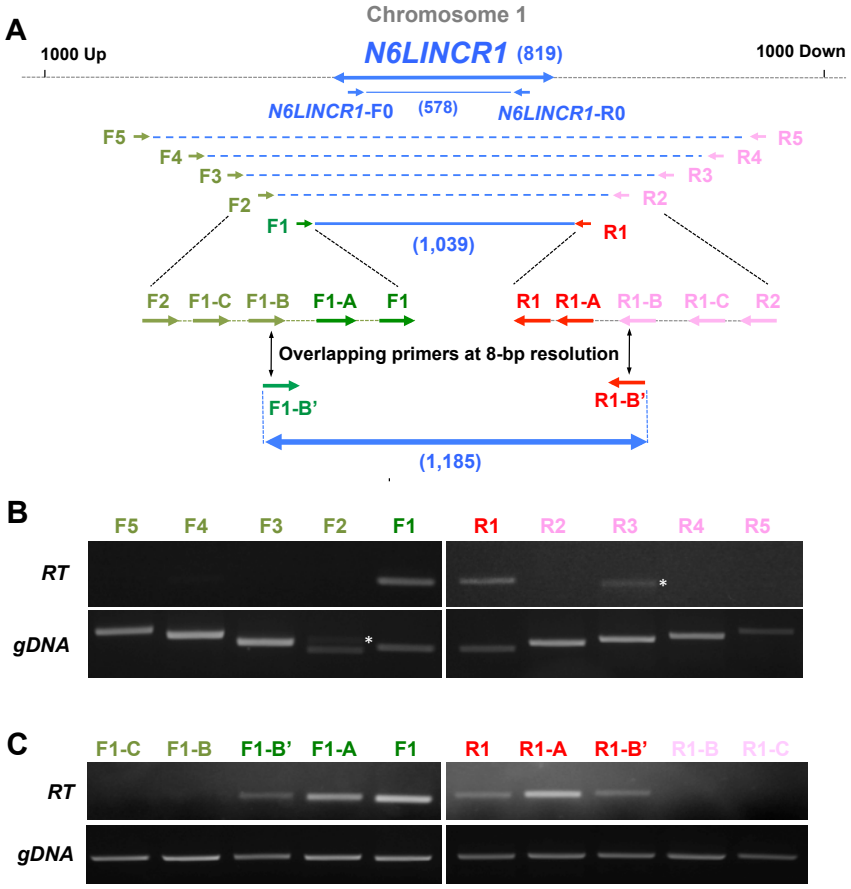
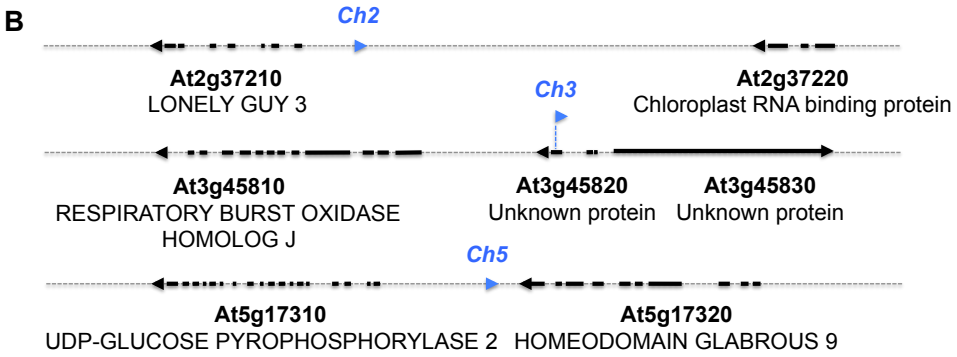


Figure S2. Identification of the 1,185-bp genomic region of *N6LINCR1* by tiling PCR. In addition to the primer set used for RT-PCR (*N6LINCR1*-F0 and *N6LINCR1*-R0) primers, five different forward (F1-F5) and reverse (R1-R5) primers, which could amplify longer *N6LINCR1* fragments than the 819-bp fragment were designed (A) and tested in RT. While all primers amplified *N6LINCR1* from genomic DNA (gDNA), only F1 and R1 amplified RT-PCR products (RT) (B). Unspecific PCR products were marked with asterisks. Then, fine-mapped primers were designed for the F1-F2 region (F1-A, F1-B, F1-C) and the R1-R2 region (R1-A, R1-B, R1-C), among which only F1-A and R1-A amplified PCR products (C). In this way, a 1,185-bp fragment of *N6LINCR1* was amplified with the F1-B' and R1-B' primers with 8-base resolution (C).

A

TAAGAGGATAATGAGCTTATAAAGCATATTATAATAATTACATGATTCATAATTTGTTGGAAATCCATT
Ch5 ATATAATTGAATCATTTGGACAAATCGCTGATAAGAAATATATTAGTAATTAAGAGAATTTCCTCTTTC
TCTTATCTCTCACATATCCTTGAGAGAGAGAGAGAGTTCACTTGTATAAACTTTTGTTTCCGGTTAGCC
GGATATGTATATCTTTCGTCTTAGTGCGATCTCAATAATTCTGGCTATGTGTCAGACTTTTGTCACCGGT
TTTGTCCGGTTCTCTCGCCGTGACGAGCTTAGATCTCCATCTTAAGCGTTTGTCGACATCTCCGCCTTCT
TCATAATTCTCTTTGTTTCCACCAAATTGTTTGAGGAATAACATCTTCATCTCTTCGTTTTGGTCACAAC
Ch3 GTACGAGCAAAGATCAAGCTACATCAATTTTGGGTGTAGCCAATGATGAAGCTTCCCCGACGATGGT
GCTAATTGGAGATTTTGAAGCAGCTCTTTAGACTTGATCCGGCAGAATATCGACATCTAGGCGCTTGC
ATTCGAGATCGACAGAAGGCACAAAGATCTTCATCGGTCACTGGCCGATTGATGGTGGCAAATCAGA
ACTGCAGTGGTGTTTGGCGATTCCAACGGTGGAGCTCGTTTGTCTCGACGCGTGGCGAAGGGGAGTA
AAACGTCCTCCACGCGTGTCAGCACTTAATCATTATTGCGTTAATGTTTTATTATATTTTCAAACGATTT
GGGTTGGACATGAGTTTAATTTTCGCTGTAATGTTGGGCCTTCTGTATTTTGGCCTTATTGGACTTTTA
Ch2 ATGGAAATGGTCGACAAAAAAAAAAAAAGAAGAAATATATTAGTAATTCTTCACATTAAAACCATAAA
ATACATTAGTAATTTGATGCAAAGATATTATTCCTTAATGGAAGTGATGGAAGTCGATTTTTAAGGCCC
AAAAAAAATAAAGCCCAAAAGACAAAAATCTAAAGTTTTGAAATTTCCTTTTCTGATTCTGAAGTTTCC
TTTTTTGTTTTGAAATTTTCTTTATATGATTCTAAAGTTTCCATTTGTTTTGTTTTTTTTTCCCTTTTTTCGA
TTCAGAGCACAACTCTATATAAGAAGCAAGACTCTTCATCAATAGTGGTGTTATCTGAACAACTGAAA
GAGAATGAGAAAC



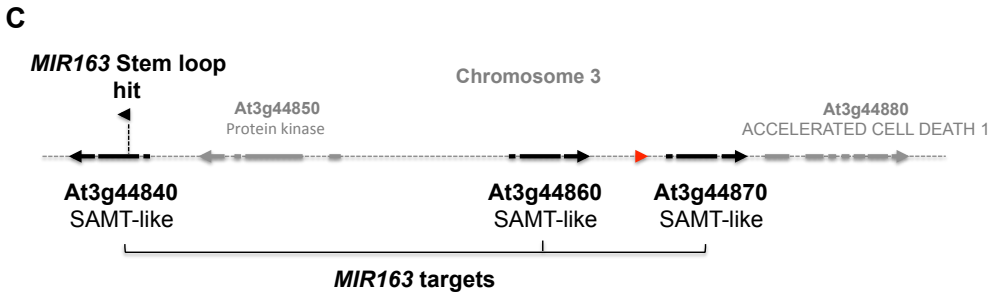


Figure S3. Analysis of the 1,185-base *N6LINCR1* sequence. (A) *N6LINCR1* (1,185 bp) sequence. The parts of sequence that matched the other chromosomes (Ch2, Ch3, Ch5) are highlighted in blue. Red lines indicate duplicated sequences. Potential triplex forming oligonucleotides (TFO) are highlighted in green. (B) and (C) Genomic regions containing the *N6LINCR1*-matched sequences (blue arrowheads indicating Ch2, Ch3, Ch5; red arrow indicating the match with the duplicated sequences).

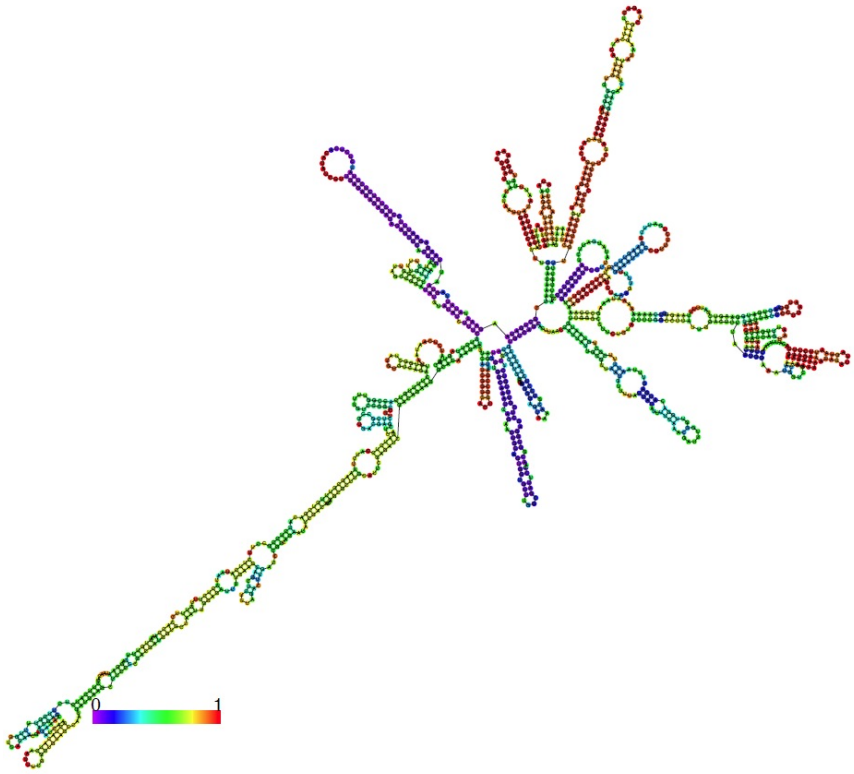


Figure S4. The structure of *N6LINCR1* predicted by RNAfold (http://rna.tbi.univie.ac.at/cgi-bin/RNAfold.cgi).

Table S1. List of primers used in this study

Primer name	Sequence
NCED6-F (#1554)	AGCTCAAACCAACGTATCCA
NCED6-R (#1555)	ACTCGTCTTTCTCTTCGTCT
ABI5-F (#1687)	ACGTCAGAGCGAGAAGTAGA
ABI5-R (#1688)	AGCAGATCAGATGGTGTTCC
DOGL4-F (#1675)	ACCACTAATGAGCAAAATGAGAAACCT
DOGL4-R (#1676)	GTTTGTGTCAGGAGTGAGTG
At3g48510-F (#1645)	TGCATAGCTAGGTACGCCAT
At3g48510-R (#1646)	TGTTCGTGCCCATAGTGACT
ACT2-F (#1872)	CCAATCGTGTGTGACAATGG
ACT2-R (#1873)	CTTCTGGGCATCTGAATCTC
InFusion-F (#1924)	CGTGGAATGCCTTCGAATAAGAGGATAATGAGCTTATAAAG
InFusion-R (#1925)	CAGGTGCTGAATTCGAAGTTTCTCATTCTCTTTCAGTTGTT
N6LINCR1-F0 (#1864)	GTTTCCGGTTAGCCGGATAT
N6LINCR1-R0 (#1865)	ACTCATGTCCAACCCAAATC
N6LINCR1-Tiling-F1 (#1874)	TGTCCTCTTTCAGCTTTTCTTGGA
N6LINCR1-Tiling-F2 (#1875)	GTCACTGTCCAATACAAACCT
N6LINCR1-Tiling-F3 (#1876)	GACAGACAACATAATGTCCCGAA
N6LINCR1-Tiling-F4 (#1877)	CACACCTCCCCATATTTG
N6LINCR1-Tiling-F5 (#1878)	CATTTGGACAAATCGCTGATAAG
N6LINCR1-Tiling-R1 (#1879)	GTTCACCACATATCTTTCATCTG
N6LINCR1-Tiling-R2 (#1880)	CTGACTAATCAAACACAGAATGC
N6LINCR1-Tiling-R3 (#1881)	AACTGAGAGCACGTGGGTTT
N6LINCR1-Tiling-R4 (#1882)	AACTCTCTCCAGGACACACT
N6LINCR1-Tiling-R5 (#1883)	GAGTTGTGCTCTGAATCGA
N6LINCR1-Tiling-F1-A (#1894)	CATGATTCATAATTTGTTGGAAATCC
N6LINCR1-Tiling-F1-B (#1893)	GAGCATAAGAGGATAATGAGCT
N6LINCR1-Tiling-F1-C (#1892)	CCATTTGGCCTTGTCTAATTTGCTGC
N6LINCR1-Tiling-R1-A (#1897)	ACCACTATTGATGAAGAGTCTTGC
N6LINCR1-Tiling-R1-B (#1896)	GGTGGGAGTTTCTCATTCTC
N6LINCR1-Tiling-R1-C (#1895)	GGTTCCACAACTAAGGAGCA
N6LINCR1-Tiling-F1B’ (#1901)	TAAGAGGATAATGAGCTTATAAAG
N6LINCR1-Tiling-R1B’ (#1902)	GTTTCTCATTCTCTTTCAGTTGTT
CYP94B3-F (#2153)	ATGGACCACCATCGTATCCA
CYP94B3-R (#2154)	TCTCCTCTCTTGACACGTGT
ALTM1-F (#2014)	TAGTAGCAGGAGGACTAGGA
ALTM1-R (#2015)	GACATGAGTCCTGTGAACTC
AT4G23680-F (#2004)	AACGTACGTGACGGAAGTTC
AT4G23680-R (#2005)	GCAACCAAGCTCTTGACGAA
STZ-F (#2169)	TCGAGGCTCTTACATCACCA
STZ-R (#2170)	ATGACTTCGTCGTCTCCGTT
ATERF2-F (#2171)	AACGAGCTGCGACTCAATGA
ATERF2-R (#2172)	CAACACGTGTCTCATCACCA
DIC2-F (#2173)	AAGGTGGGATTGCCTCTGTA
DIC2-R (#2174)	TCTCGAAGCAGCTTCCTAAC
MYB15-F (#2179)	ACGTATGGCACACTCACTTG
MYB15-R (#2180)	ACGTATGGCACACTCACTTG
PUB22-F (#2220)	CTCATGTCCAGTCACCAAAC
PUB22-R (#2221)	GAACTCAGCTCTTCCTTCTG
At3g10930-F (#2163)	ATGCTCTTCCGAAAGGGAGT
At3g10930-R (#2164)	GATGAAACGATACAAGATGATAGTC
LOB41-F (#2175)	AAGCGCAAGCTAACGCAACT
LOB41-R (#2176)	AGCTCAGTCTTACACGTGCT

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